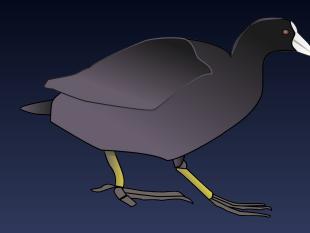
June 2010 Chicago



Ligand fitting and Validation with Coot

(Paul Emsley) (University of Oxford)

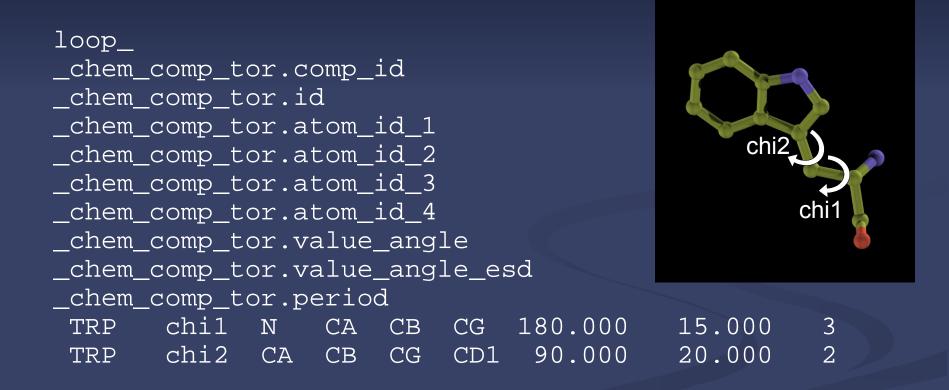
Bernhard Lohkamp Karolinska Institutet

Fitting Ligands

REFMAC Monomer Library chem_comp_bond

<pre>loop_ _chem_comp_bond.comp_id _chem_comp_bond.atom_id_1 _chem_comp_bond.atom_id_2 _chem_comp_bond.type _chem_comp_bond.value_dist</pre>					
_chem_comp_bond.value_dist_esd					
ALA	N	H	single	0.860	0.020
ALA	N	CA	single	1.458	0.019
ALA	CA	HA	single	0.980	0.020
ALA	CA	CB	single	1.521	0.033
ALA	CB	HB1	single	0.960	0.020
ALA	CB	HB2	single	0.960	0.020
ALA	CB	HB3	single	0.960	0.020
ALA	CA	С	single	1.525	0.021
ALA	С	0	double	1.231	0.020

REFMAC Monomer Library chem_comp_tor

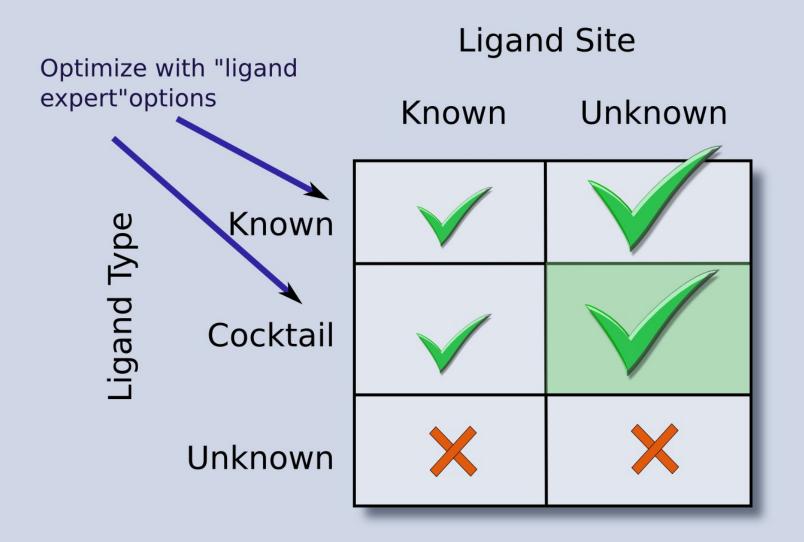


Ligand Fitting

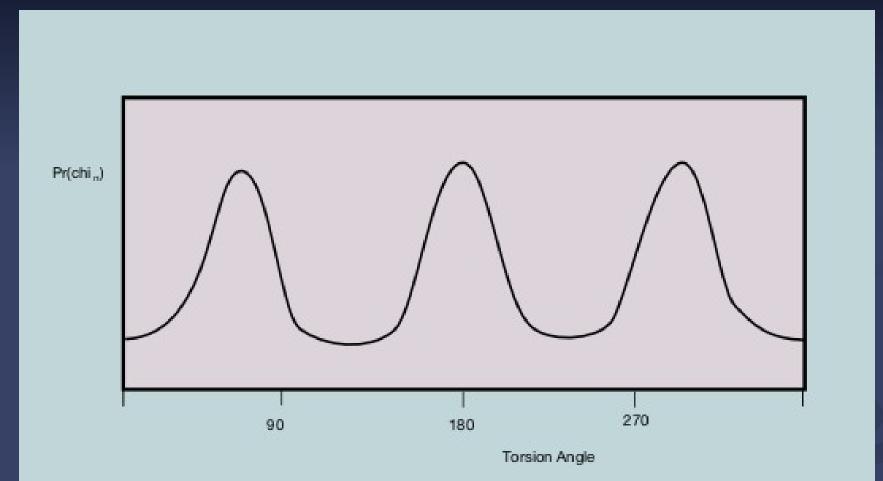
c.f. Oldfield (2001) *Acta Cryst. D* X-LIGAND

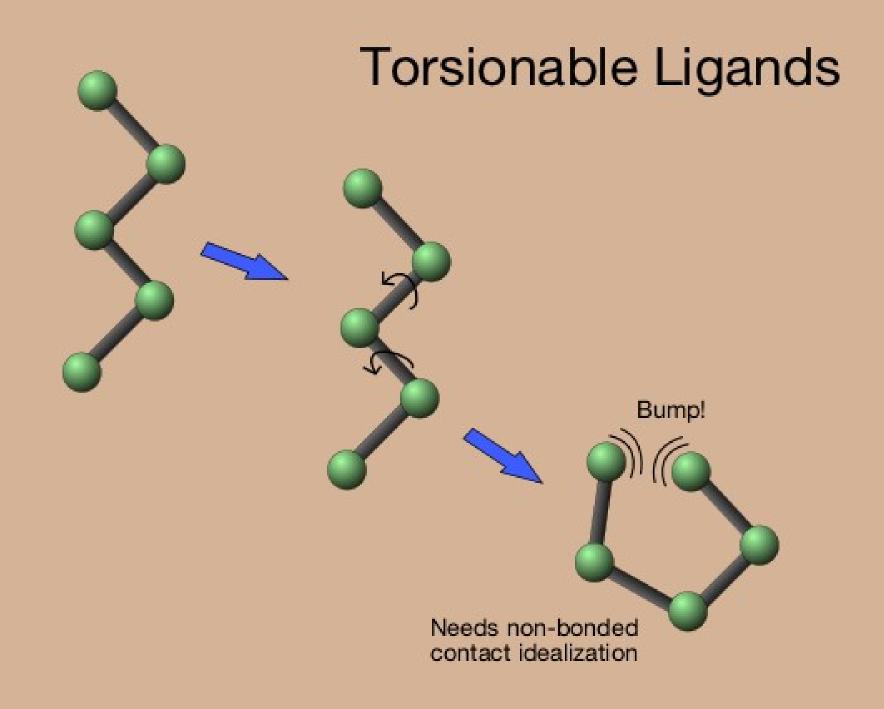
Somewhat different torsion search algorithm

Build in crystal-space



Ligand Torsionable Angle Probability from CIF file





Crystal Space

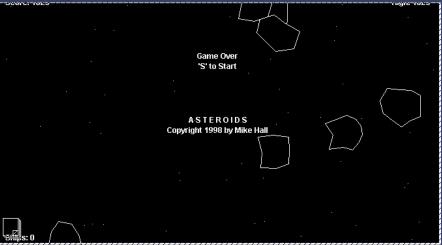
Build in "crystal space"

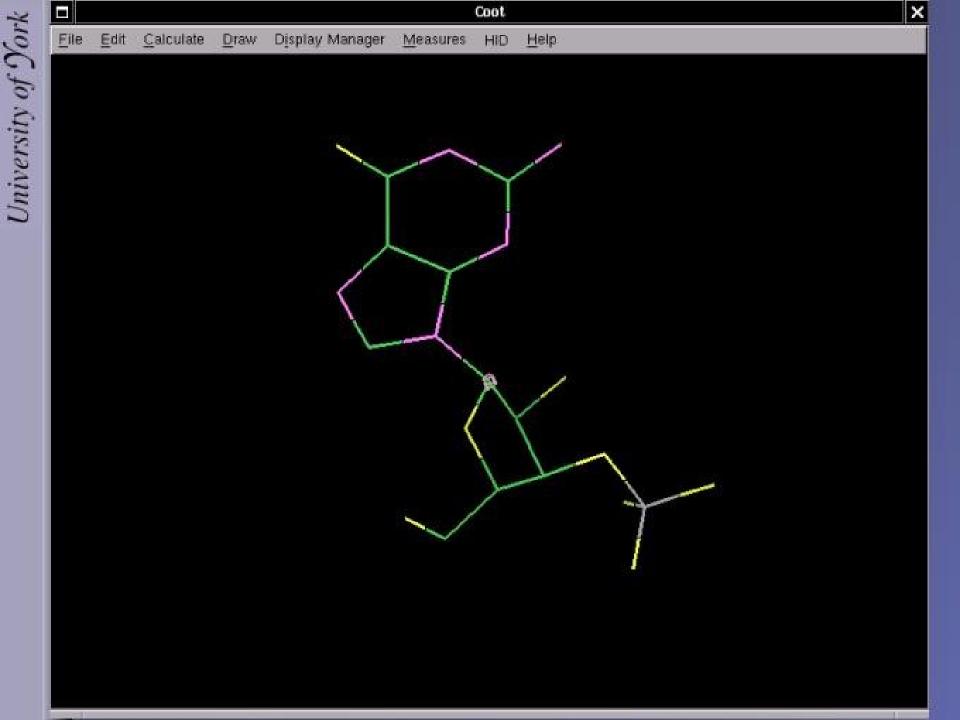
- Like real-space, but wrapped by crystal symmetry
 - Like "Asteroids"
- Assures only one real-space representation of map features
 - Build everything only once,
 - No symmetry clashing
- However, more difficult to calculate real space geometries
 -such as bonds, torsions

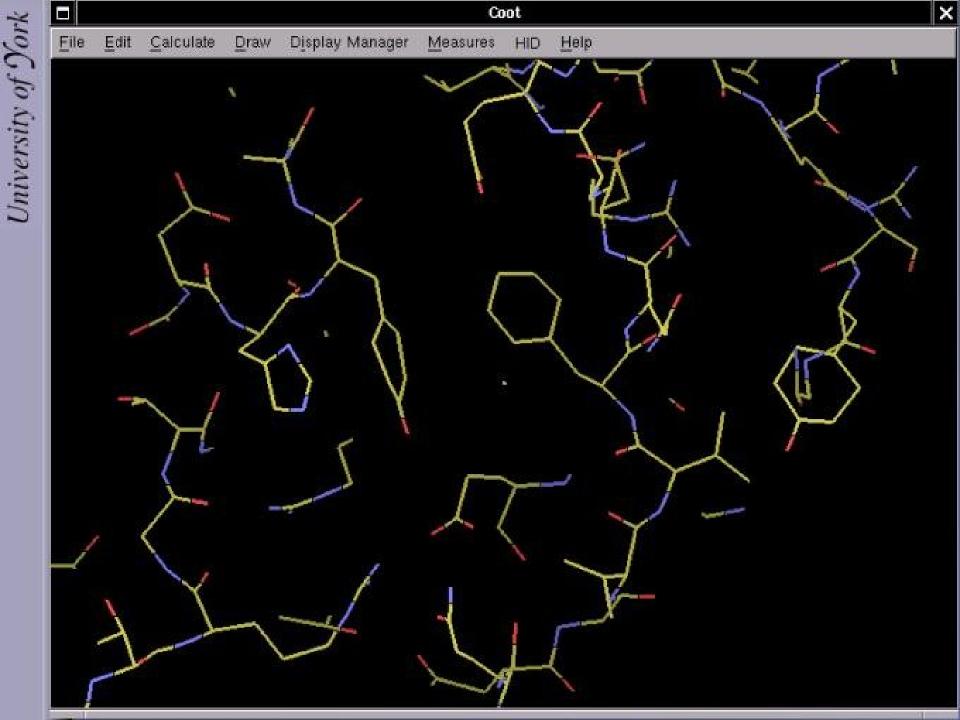
Crystal Space

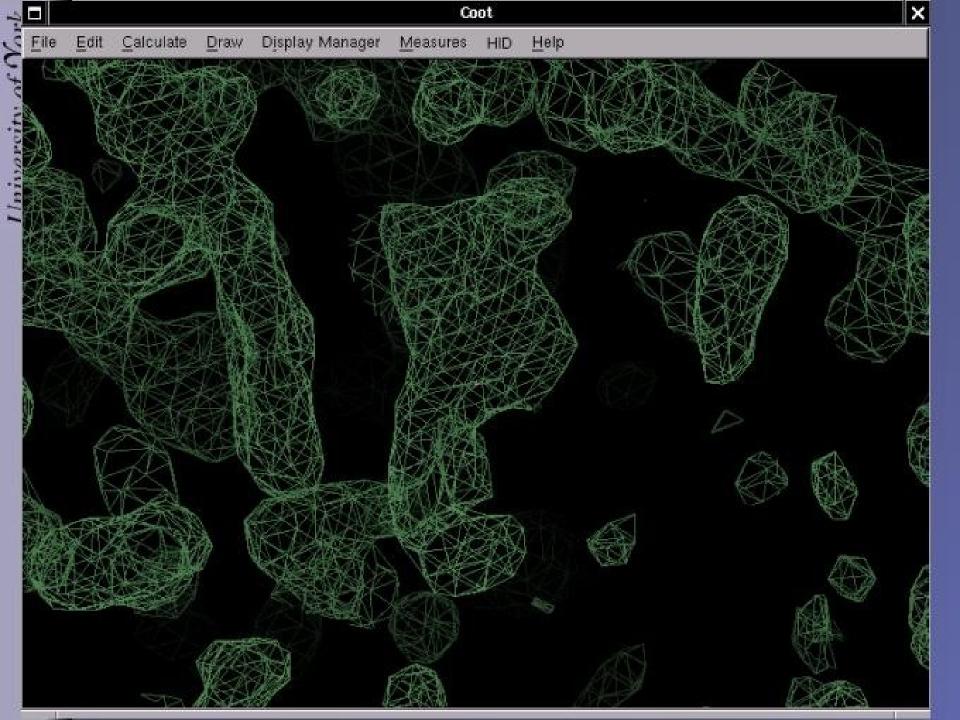
Build in "crystal space"

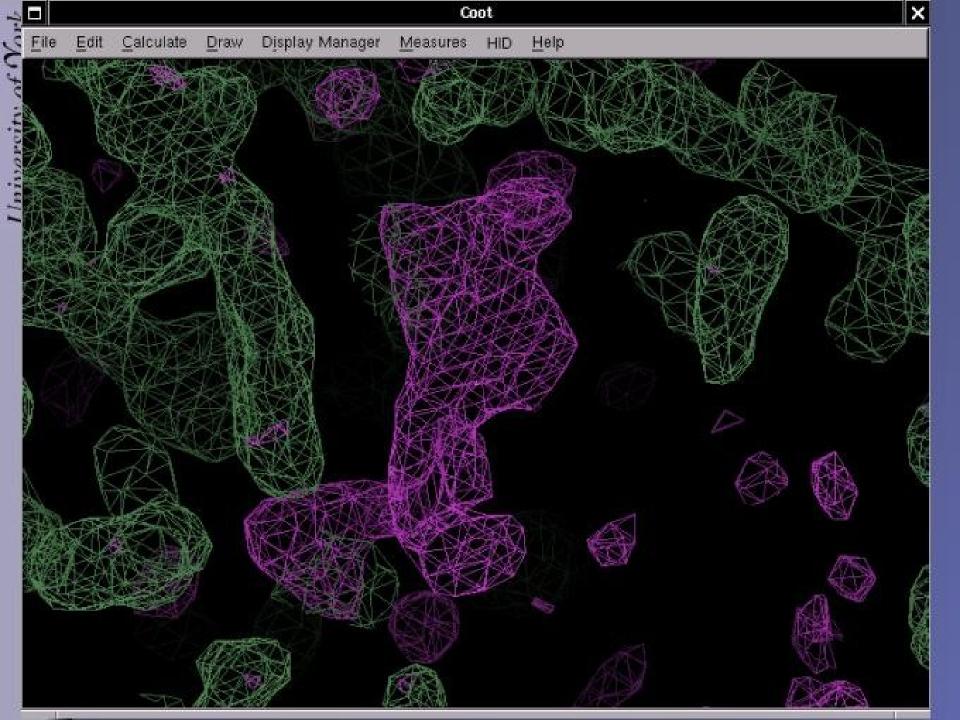
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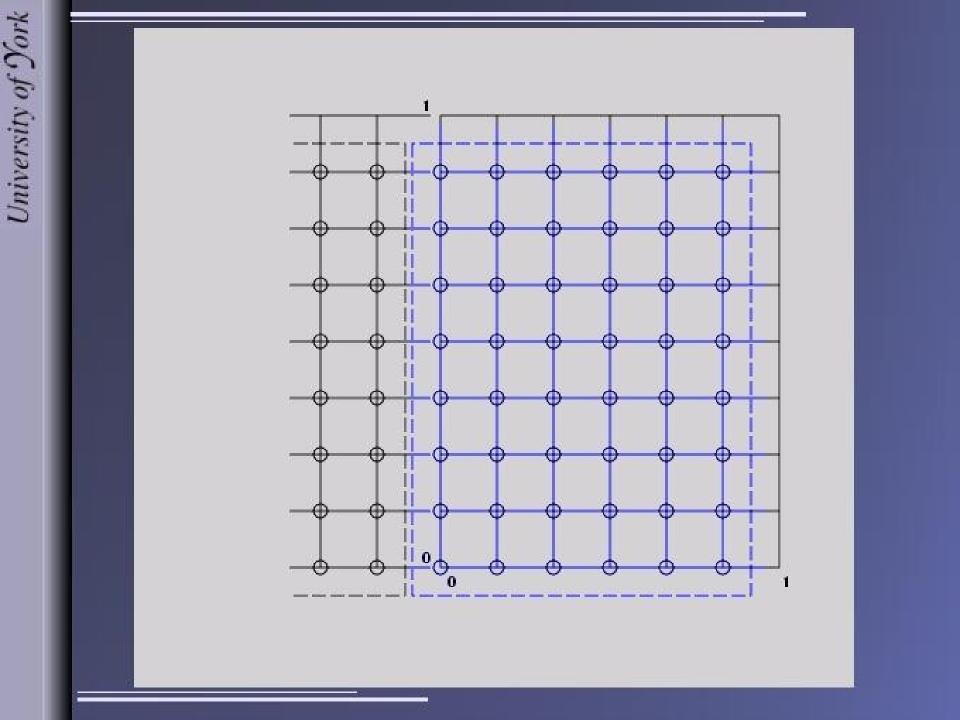


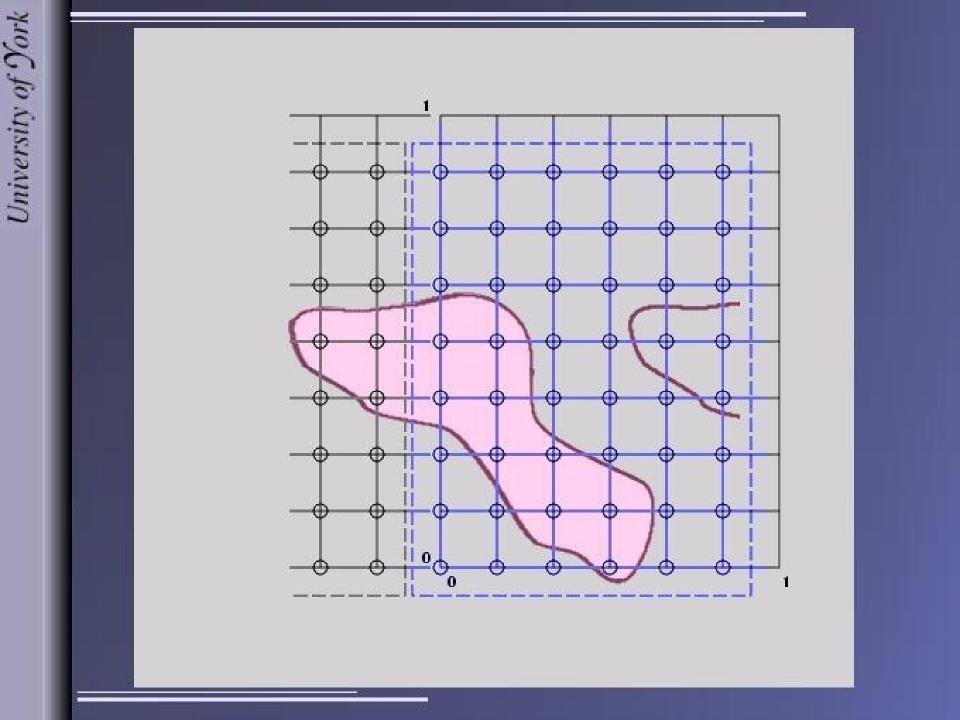


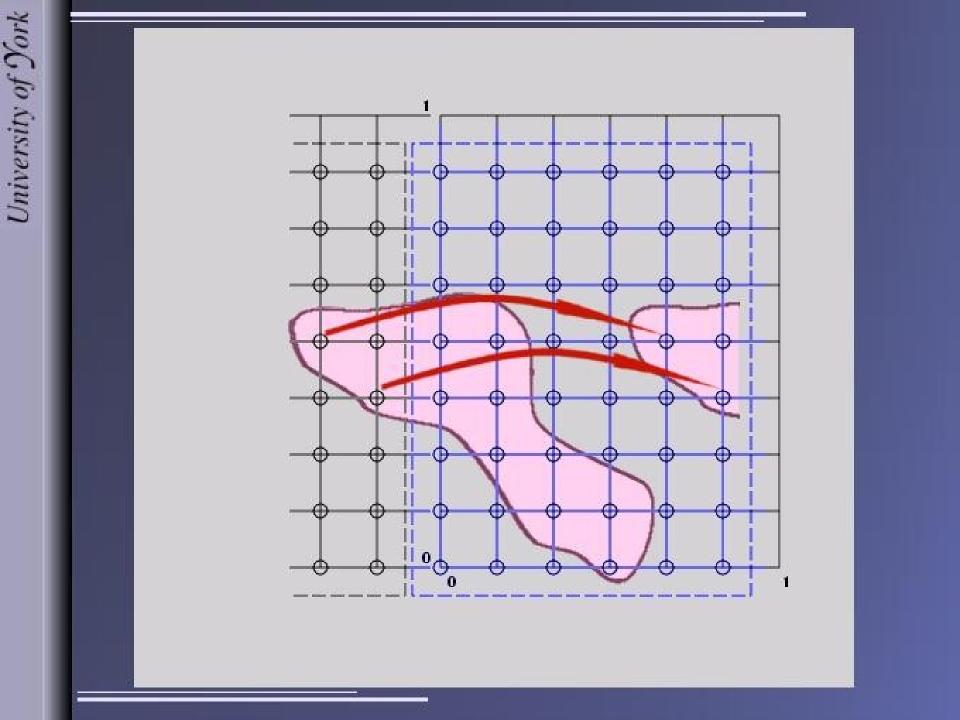
Clipper Map Mapping

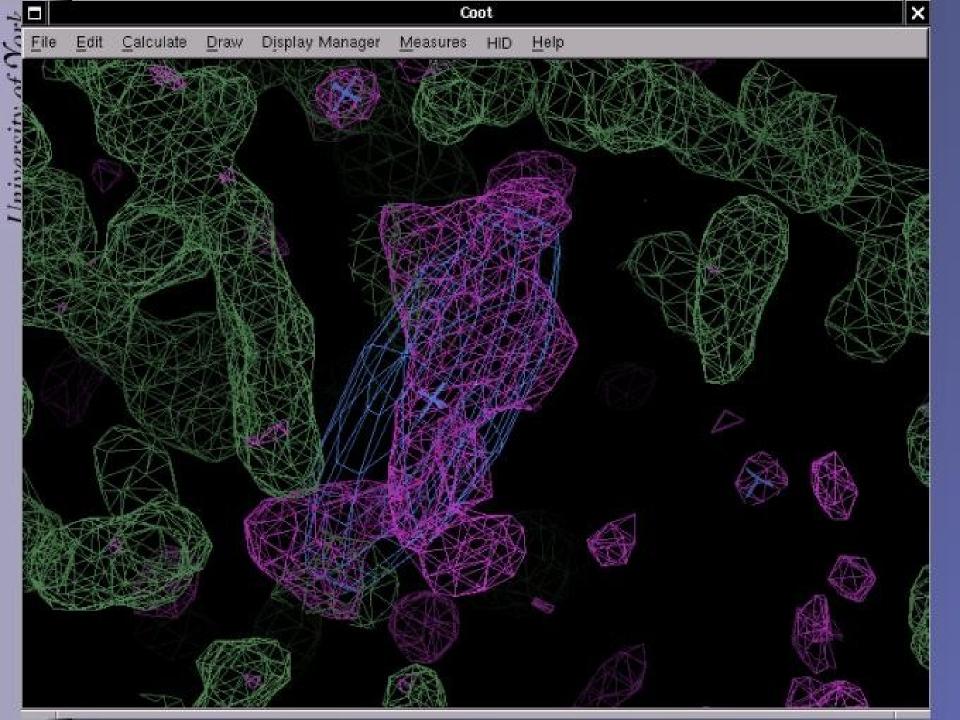
Clipper maps

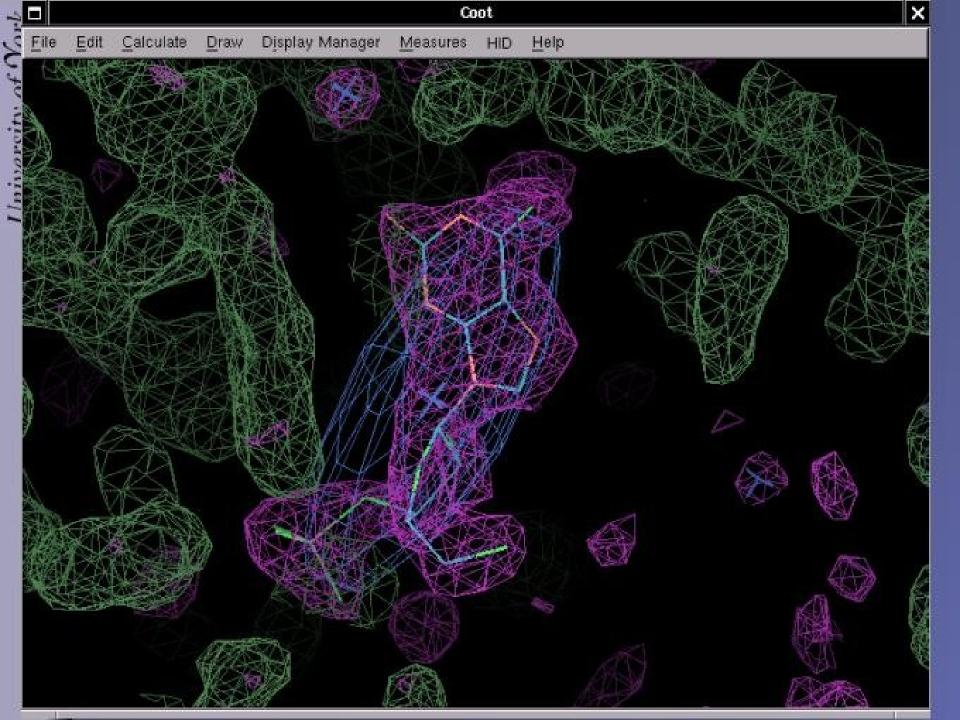
- Appear to be "infinite"
- Density value can be queried anywhere in space

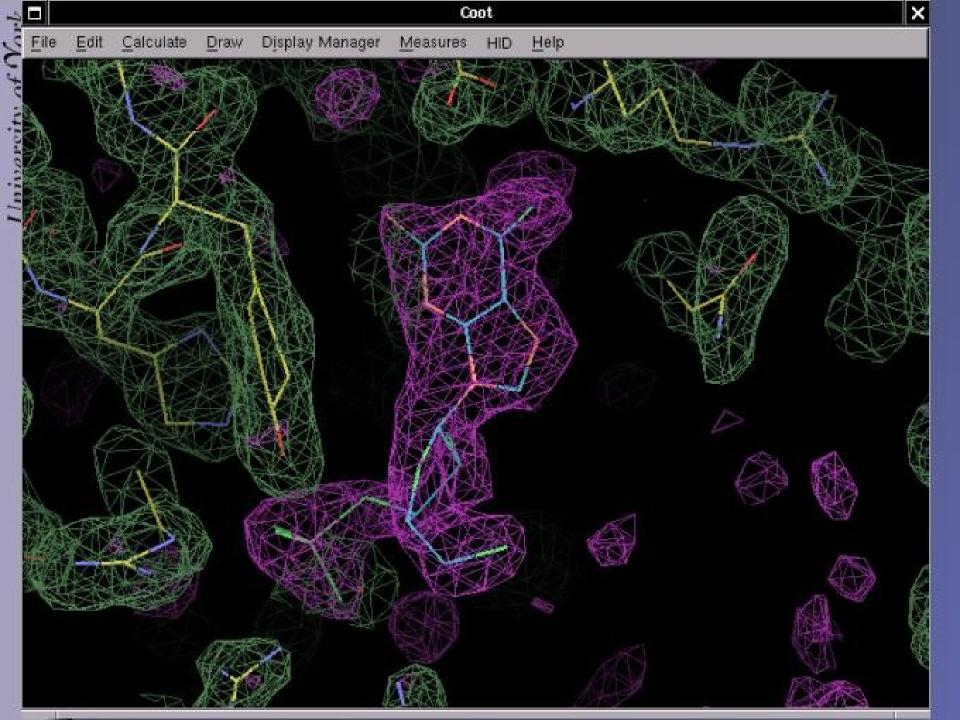












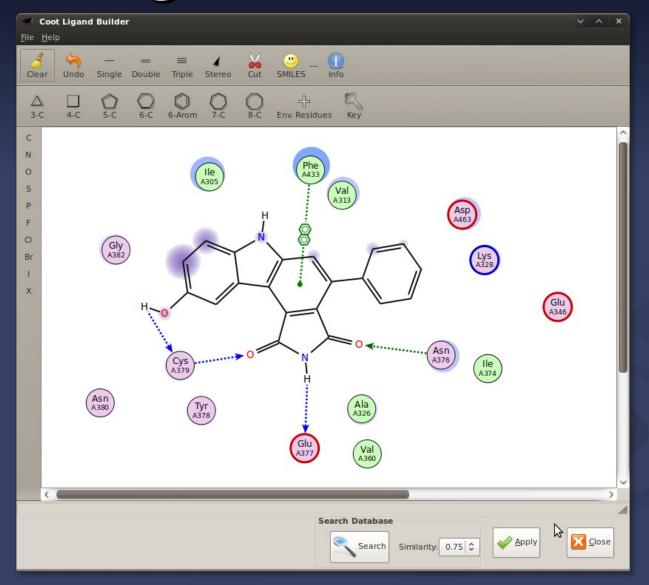
Conformation Idealization

- Each conformer is passed through the "Regularization" function of Coot
 - Non-bonded terms included
- Better to have hydrogen atoms on the model
- Slows things down a good deal...
 - May not be the best method to explore conformational variability for many rotatable bonds

Ligand Overlay

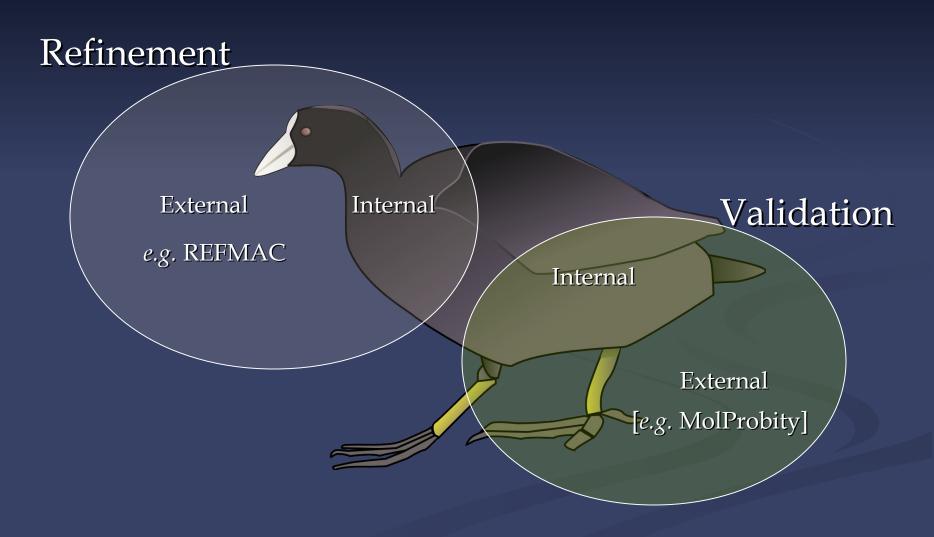
- Algorithm and Code by Eugene Krissinel
- Tries to overlay different ligands/monomers by graph matching
- Useful for "database" ligands where atom names are not selected by hand
- Has been used as the basis of the function which "mutates" residues to alternative monomer types
 e.g. phosphorylation

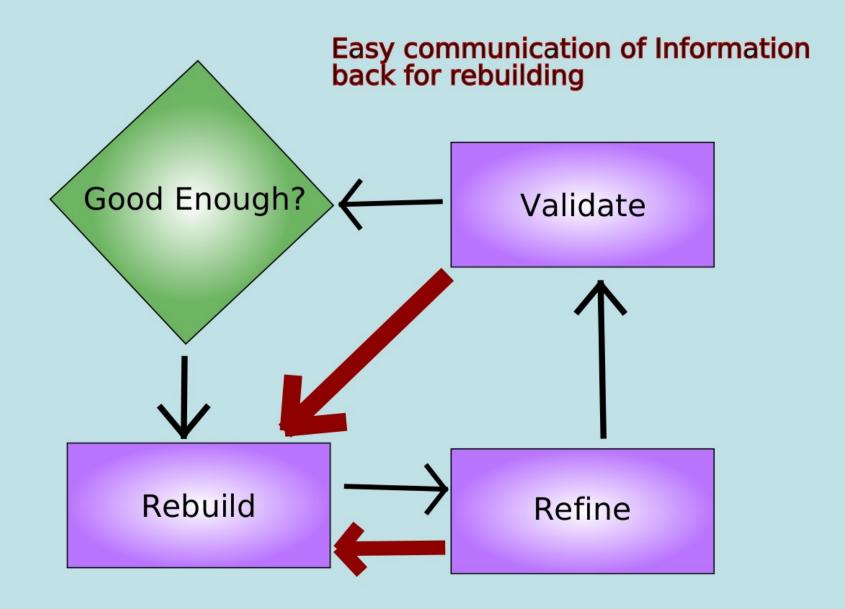
New ligand view and builder



Coming soon...

Feature Integration





Validation...

What is Validation?

- Comparison of Various aspects of the model with pre-conceived notions of "good quality"
 - Includes unrestrained and restrained criteria
 - Many aspects of validation overlap with refinement and model-building

Why Validate?

- Model-building is error-prone
 - (although automated methods seem to do better)
- Someone else did the model-building
 The model was built several years ago

 and the notion of "good quality" has changed

 Deposition requires validation

Observations to Parameters Ratio

- Some typical numbers
 - to 2Å, 22000 reflections
 - 200 residues x 10 (atoms/residue) x 4 params/atom
 - -> about 2.6
- **To 3Å**:
 - Ratio is about 1:1
- As statisticians, we prefer our models to be parsimonious

Depending on solvent content and the manner in which NCS is handled

A "good" model

Makes statistical sense

- The reciprocal space representation agrees tolerably well with the observations (R-factor)
- No meaningful difference map peaks
- Makes Chemical sense
 - Model Geometry is consistent with the restraints
 - Ramachandran Plot has less than 1% outliers
 - A good clashscore
- Makes Biological sense
 - Residues in chemically sensible environment
 - Is consistent (on the whole) with external biochemistry observations (active site residues)

Quick Bayes

- Bayes Eq:
- Pr(model | data) ~ Pr(data | model) * Pr(model)
 Pr(data | model) is also called the Likelihood, L(model | data)

Validation Tools - Pr(model)

Ramachandran Plot Kleywegt Plot (NCS differences) Geometry Analysis Peptide ω Analysis Temperature Factor Analysis **Rotamer** Analysis [Clashes]

Rotamers

- Side-chains have certain preferred combinations of torsions round their rotatable bonds
- An analysis (batched around the staggered conformations) will give rotamer occurrence

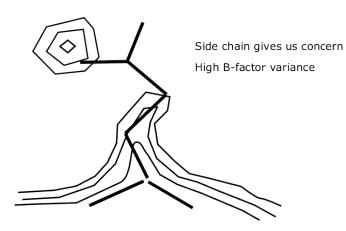
Validation Tools -Pr(data|model)

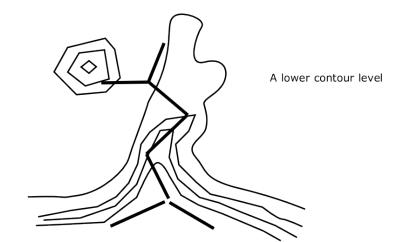
Density Fit Analysis
Difference Map Peaks

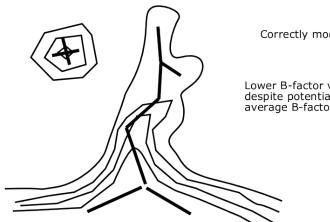
Variance analysis at Water Positions

Unmodelled blobs

B-factor variance





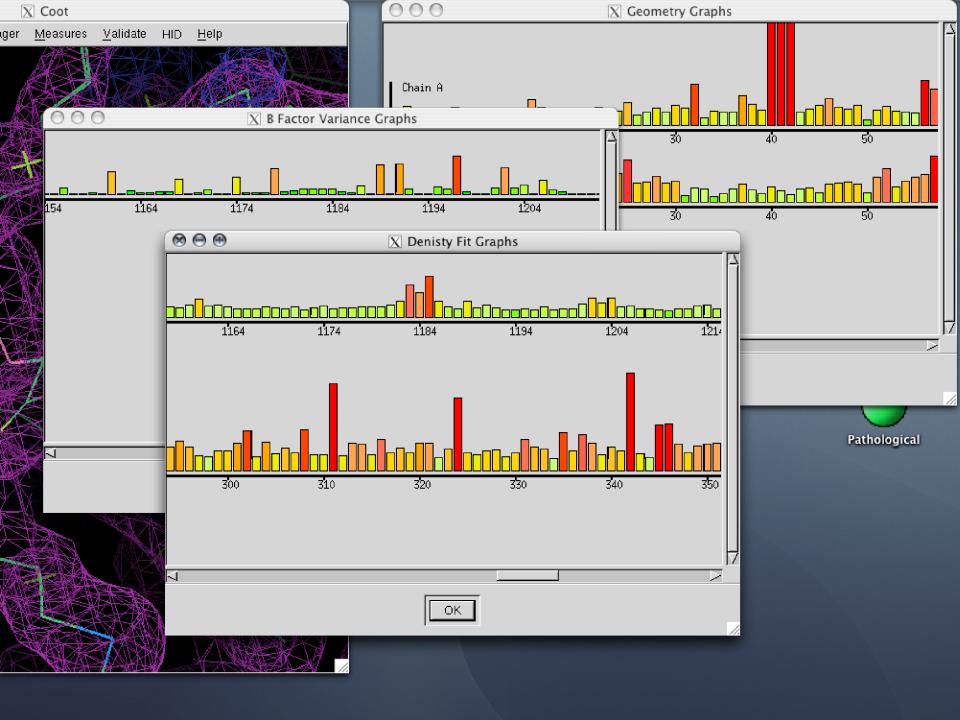


Correctly modelled side-chain

Lower B-factor variance despite potentially higher average B-factor

Chiral Volume Analysis

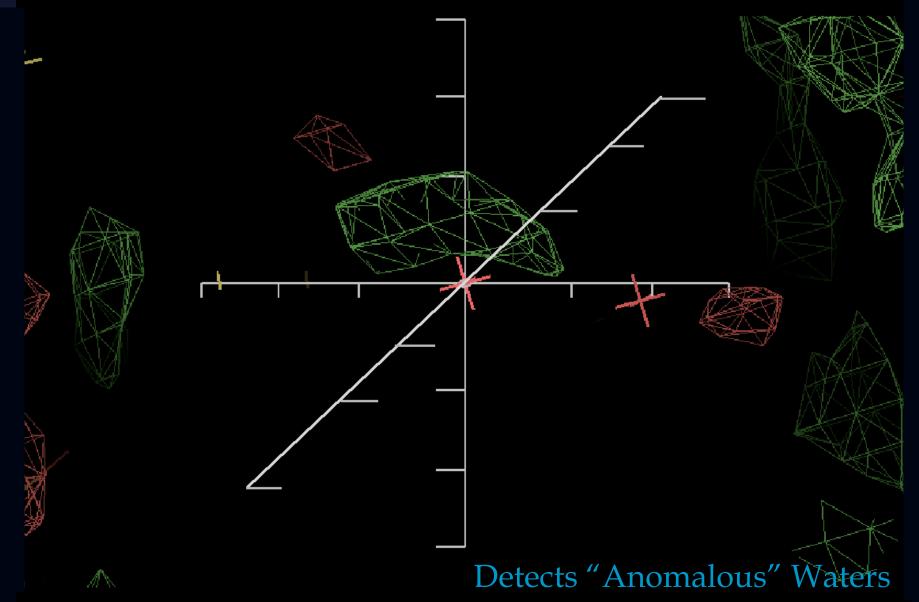
- Based on data in the Refmac dictionary
- ...was needed because it was possible with Coot to accidentally invert Chiral centres
 - *e.g.* $C\alpha s$, $C\beta$ (THR)
 - (Easily corrected with the Mutate & Autofit tool)
 - These days we have chiral volume restraints

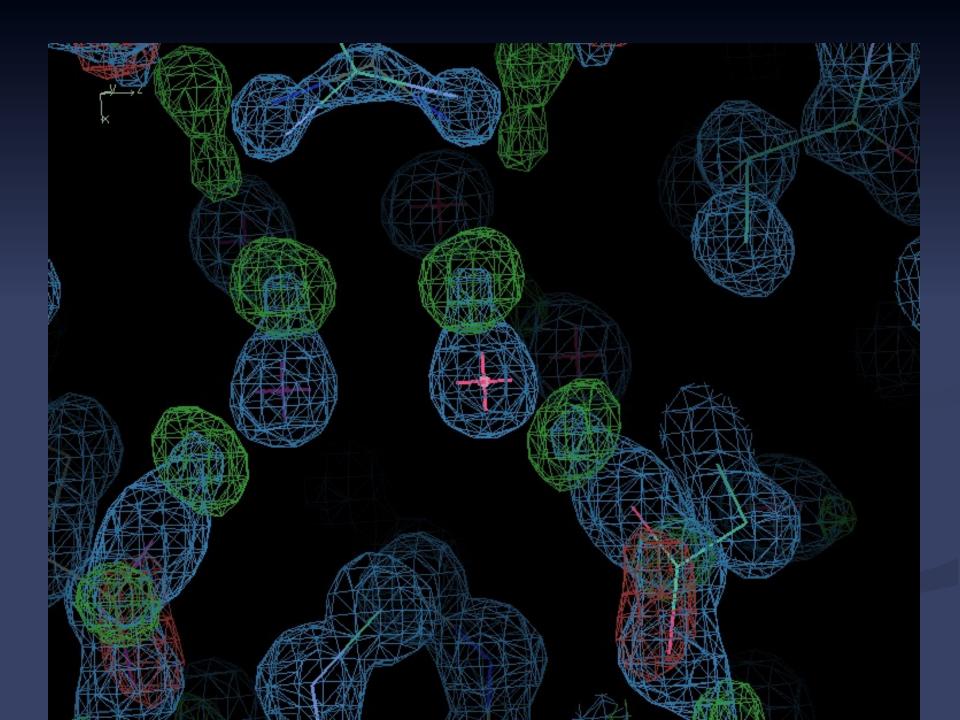


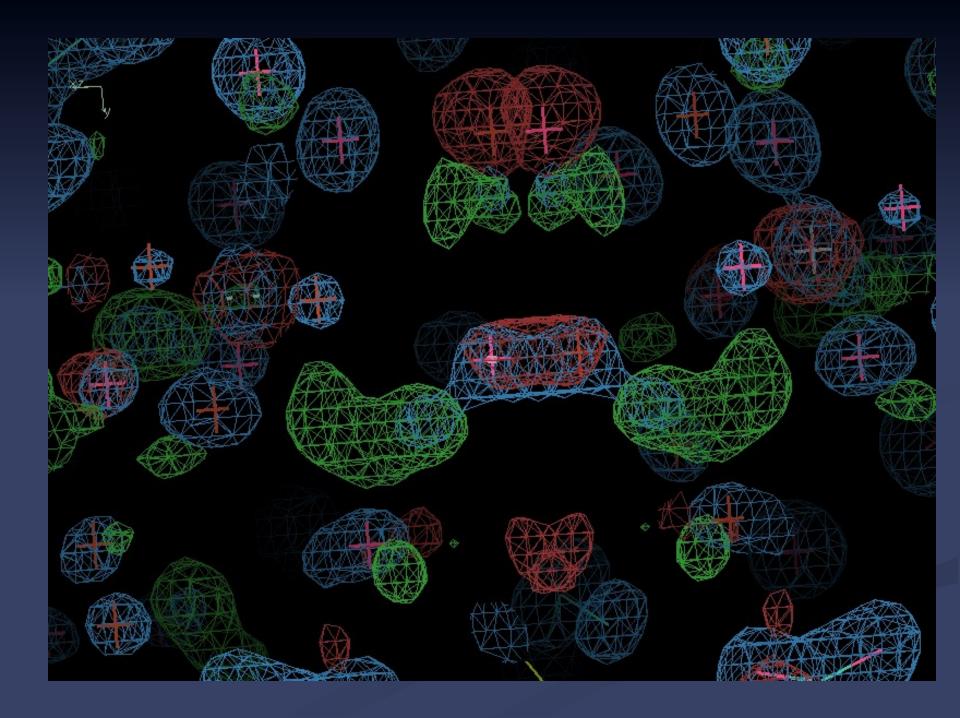
Check/Delete Waters

$\Theta \Theta \Theta$	X Chec	king Waters	:	
Molecule Number: 2aulemsley/data/pad/pad15_8.pdb			_	
Action: Generate a List 💷				
with B factor greater than: 80.00 A^2				E Active
with map sigma level less than: 1.00 electrons/A3				F Active
with closest contact less than: 2.30 A				F Active
with closest contact more than: 3.50 A			F Active	
Ignore Partial Occupancy close contacts				
☐ Ignore Waters with Zero Occupancy				
For Waters that match:				
Any Criterion				
🕹 All Criteria				
ок			Cancel	

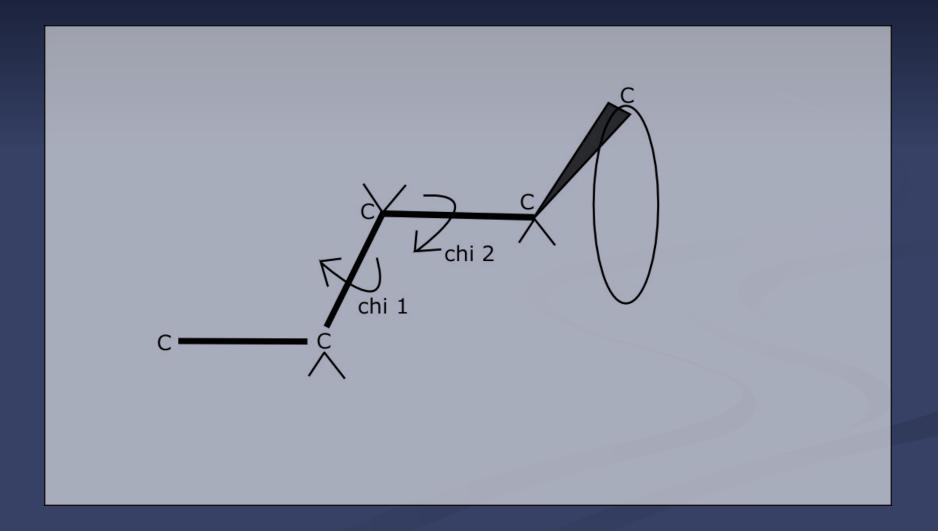
Difference Map Sampling

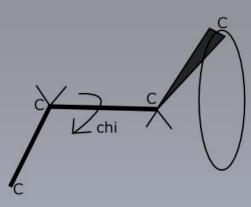






Torsion-based Validation

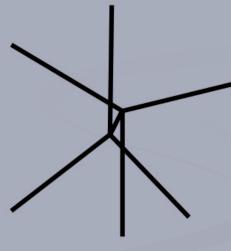




In principle, there is free rotation

In practice, staggered is energetically more favoured



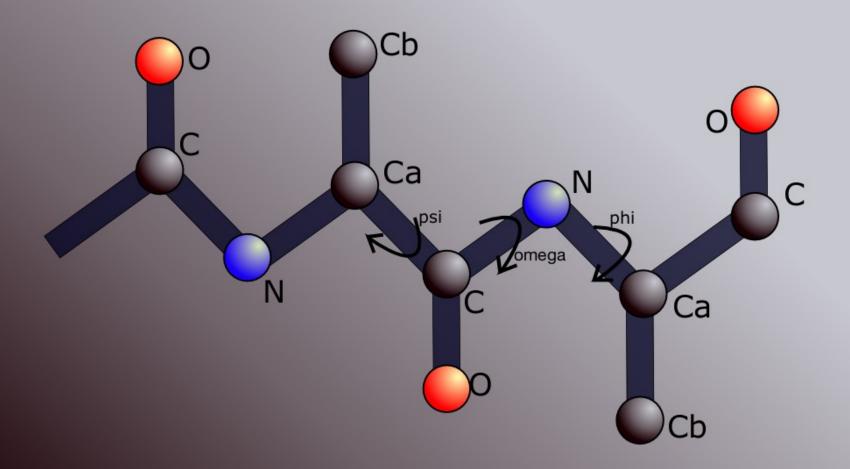


Eclipsed

Staggered

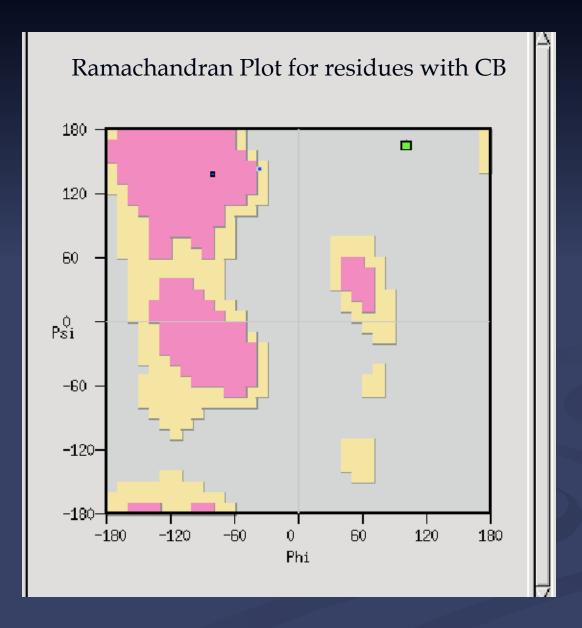
Most favoured staggering angles 60, 180, -60 degrees

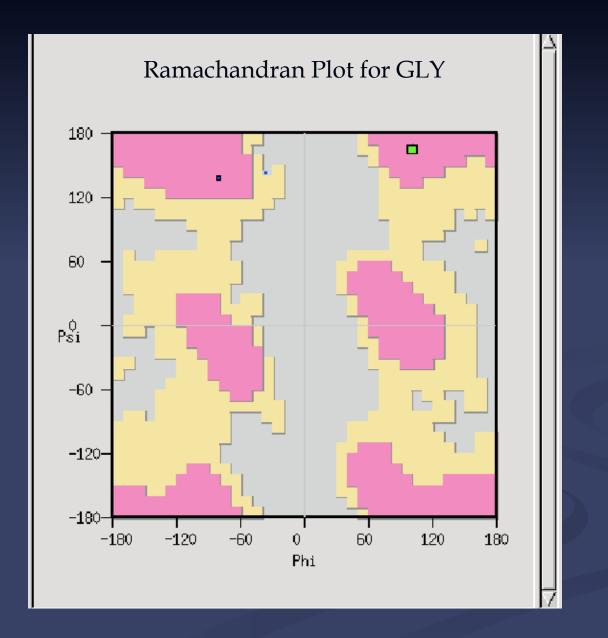
Peptide Torsion Angles

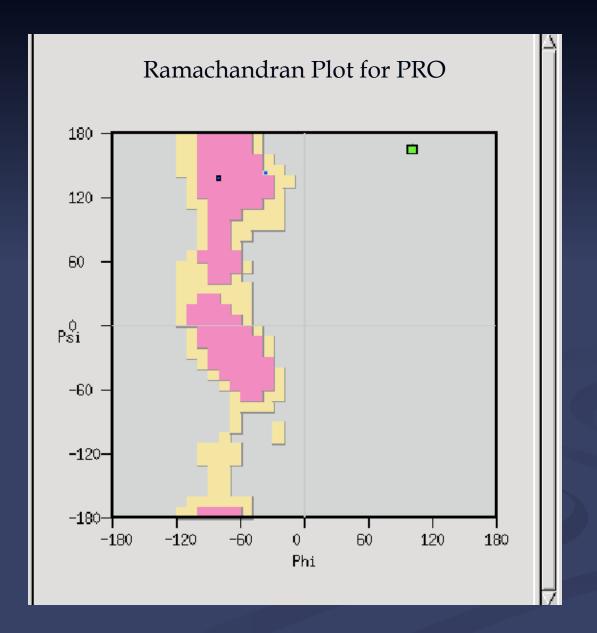


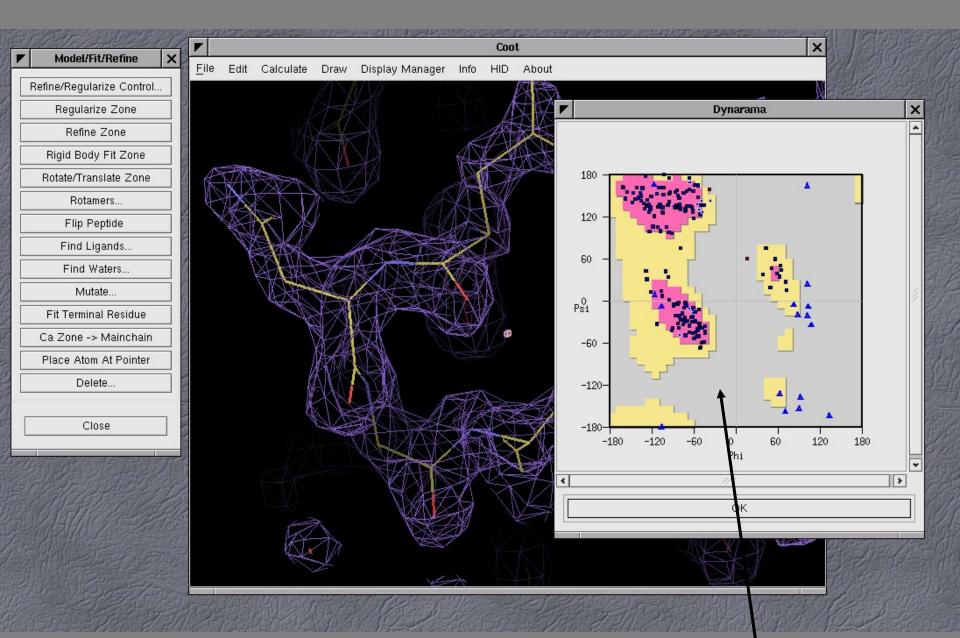
Peptide w

- Needed to check the planarity of the peptide link
 - At low resolutions it is possible to give the protein lots of (too much) freedom to optimize the fit to the density
- Can accidentally create CIS peptides
 - When discovered they are easily reconverted using the CIS<->TRANS peptide tool
- Less accidents happen when peptide plane restraints are applied



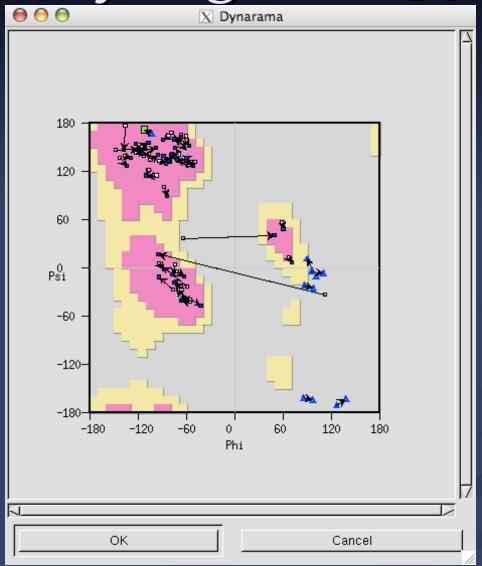






Top500-based distribution

Kleywegt Plots[*]

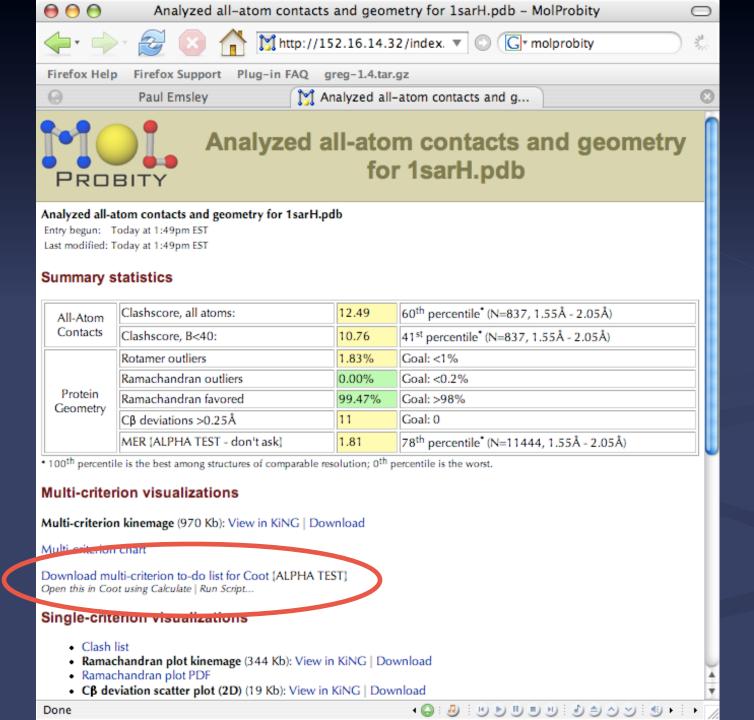


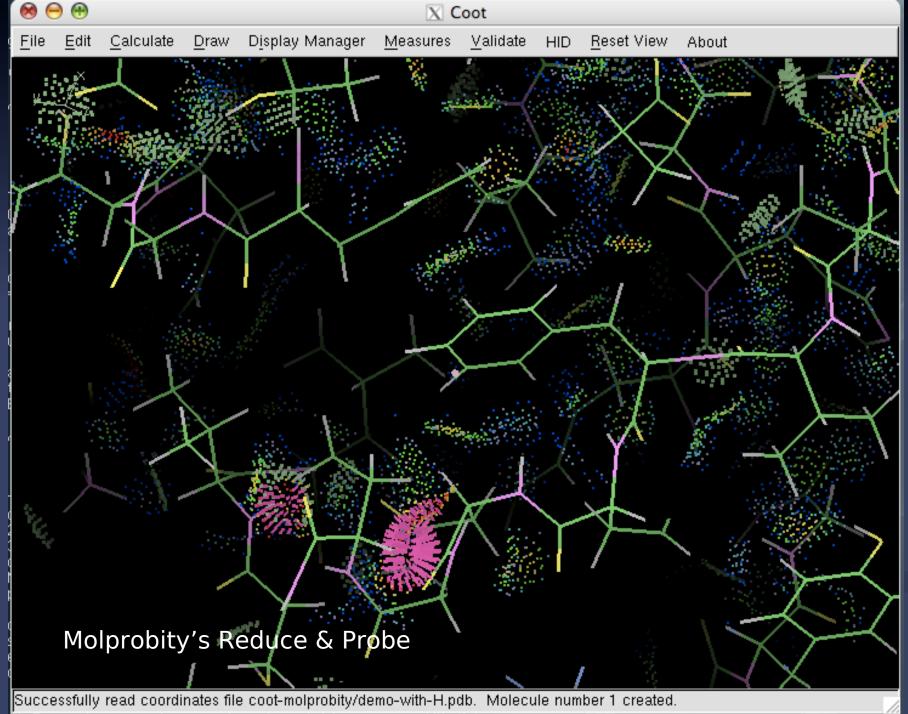
[*] Named by George Sheldrick

More Validation Pr(model)

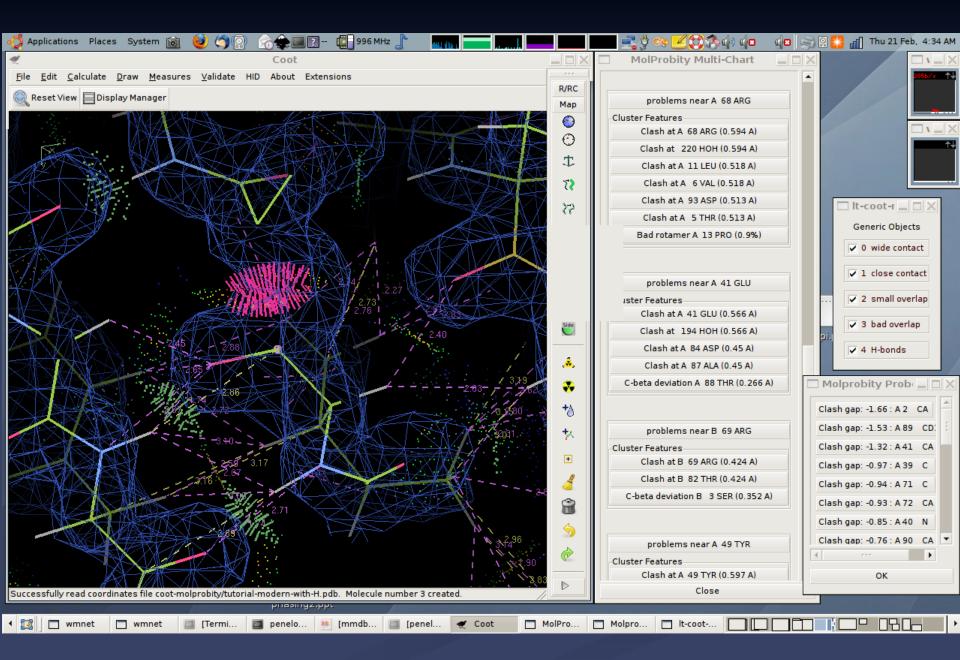
Coot has interface to Molprobity

- (Molprobity is the widely regarded as the best model validation suite)
- Uses identical Ramachandran plot
- Uses identical Rotamer library
- Coot reads probe dots directly





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Other Programs

Moprobity Suite
 molprobity.biochem.duke.edu
 WHATCHECK
 VERIFY-3D

Acknowledgements

- Paul Emsley
- Kevin Cowtan
- Eleanor Dodson
- Keith Wilson

http://www.biop.ox.ac.uk/coot/ or Google: Coot

or for WinCoot

http://www.ysbl.ac.uk/~lohkamp/coot

- Libraries, dictionaries
 - Alexei Vagin, Eugene Krissinel, Stuart McNicholas
 - Dunbrack, Richardsons
- Coot Builders and Testers
 - William Scott, Ezra Peisach
 - York YSBL, Dundee, Glasgow (early adopters)
 - Coot Mailing List subscribers