

Ligand fitting and Validation with Coot

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Fitting Ligands

REFMAC Monomer Library chem_comp_bond

```
loop_
_chem_comp_bond.comp_id
_chem_comp_bond.atom_id_1
_chem_comp_bond.atom_id_2
_chem_comp_bond.type
_chem_comp_bond.value_dist
_chem_comp_bond.value_dist_esd
                        single
                                              0.020
 ALA
                                    0.860
          \mathbf{N}
                Η
                        single
                                              0.019
ALA
          N
                CA
                                    1.458
 ALA
          CA
                        single
                                    0.980
                                              0.020
               HA
                        single
                                              0.033
 ALA
          CA
                CB
                                    1.521
                        single
          CB
               HB1
                                    0.960
                                              0.020
 ALA
                        single
 ALA
          CB
               HB2
                                    0.960
                                              0.020
 ALA
          CB
               HB3
                        single
                                    0.960
                                              0.020
                        single
                                    1.525
                                              0.021
 ALA
          CA
                C
                        double
                                              0.020
                                    1.231
 ALA
          C
                \bigcirc
```

REFMAC Monomer Library chem_comp_tor

```
loop
_chem_comp_tor.comp_id
_chem_comp_tor.id
_chem_comp_tor.atom_id_1
_chem_comp_tor.atom_id_2
_chem_comp_tor.atom_id_3
_chem_comp_tor.atom_id_4
_chem_comp_tor.value_angle
_chem_comp_tor.value_angle_esd
_chem_comp_tor.period
      chil N
                        CG 180.000
                                      15.000
TRP
                    CB
                                               3
                CA
TRP chi2 CA
                             90.000
                                      20.000
                CB CG
                        CD1
```

Ligand Fitting

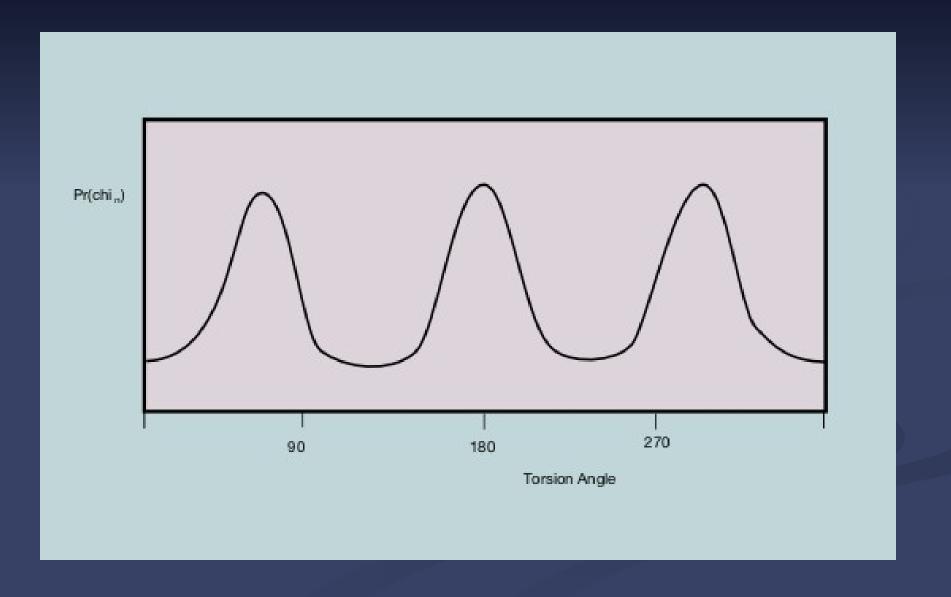
c.f. Oldfield (2001) Acta Cryst. D x-LIGAND

Somewhat different torsion search algorithm

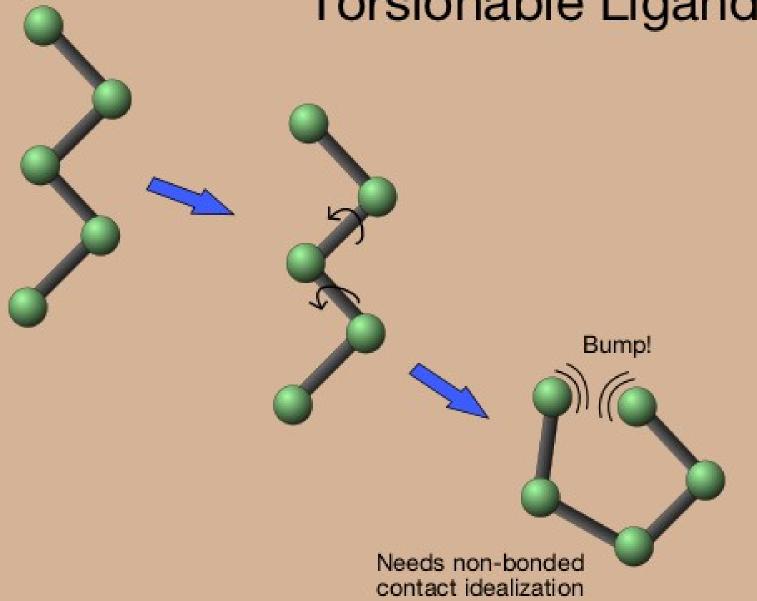
Build in crystal-space

Ligand Site Optimize with "ligand expert"options Known Unknown Known Ligand Type Cocktail Unknown

Ligand Torsionable Angle Probability from CIF file

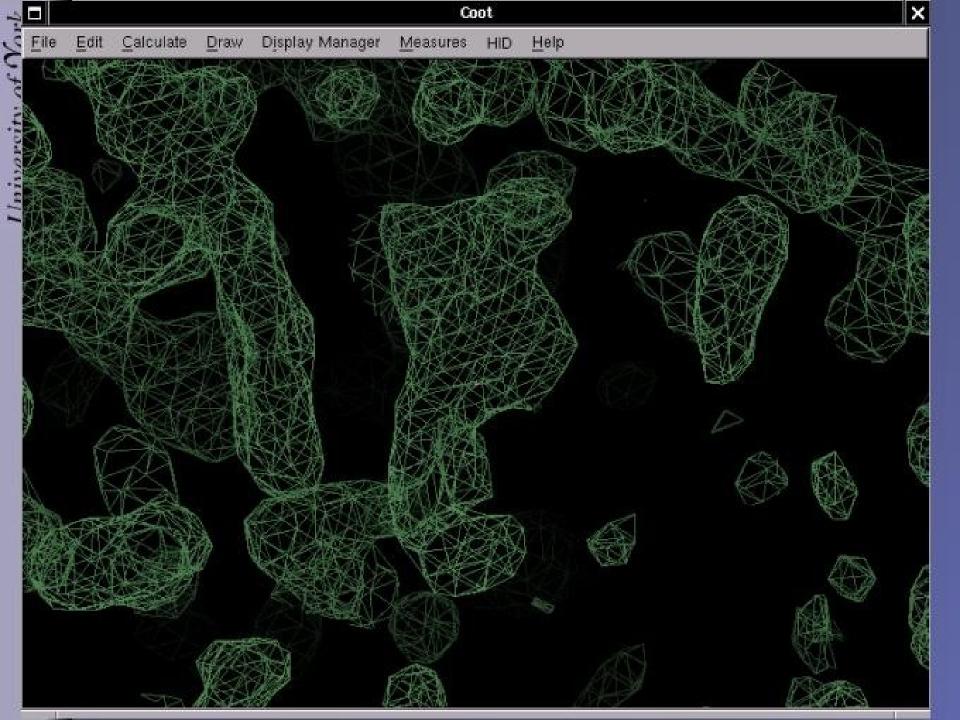


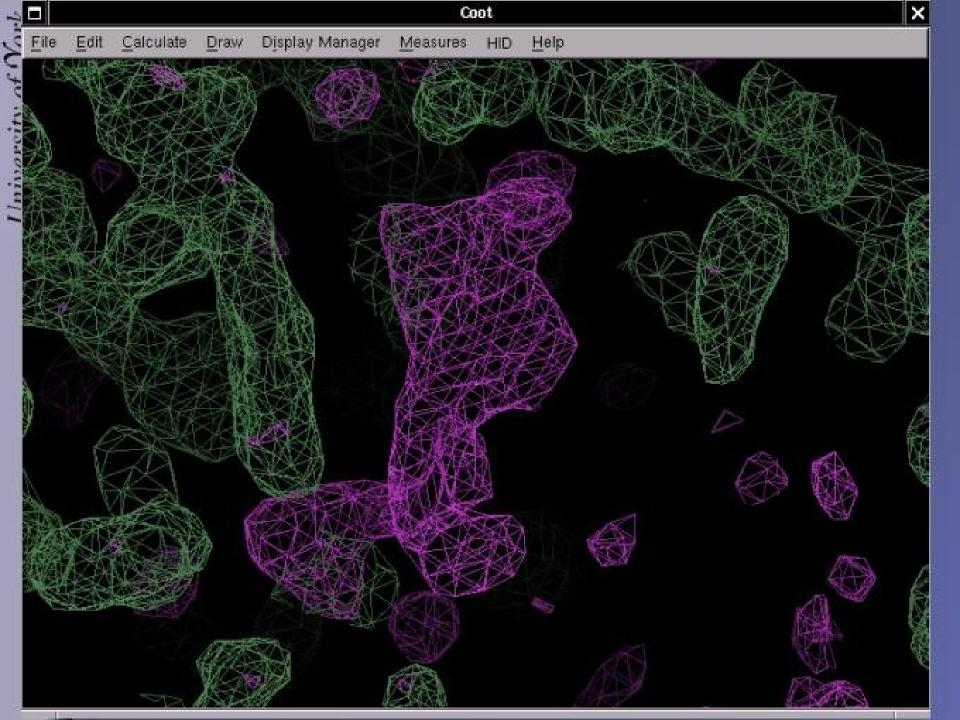
Torsionable Ligands



Crystal Space

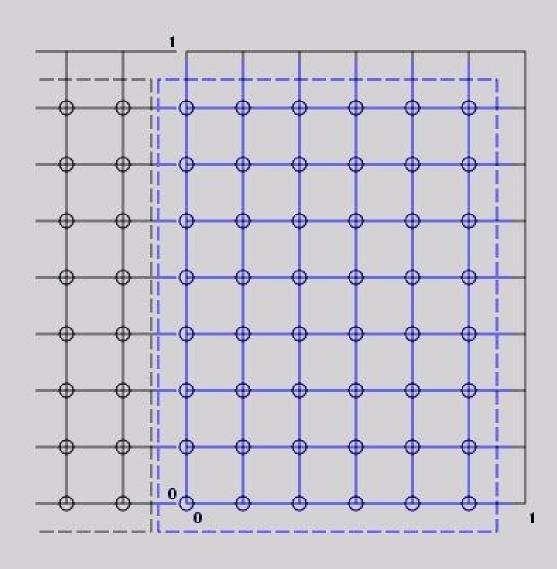
- Build in "crystal space"
 - Like real-space, but wrapped by crystal symmetry
 - Like "Asteroids"
 - Assures only one real-space representation of map features
 - Build everything only once,
 - No symmetry clashing
 - However, more difficult to calculate real space geometries
 - ...such as bonds, torsions

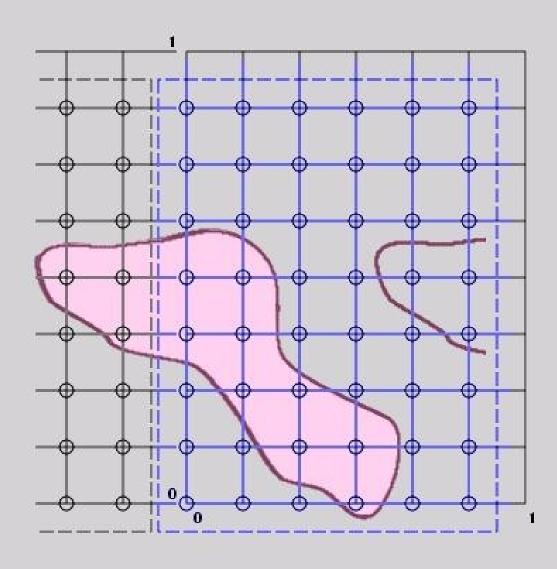


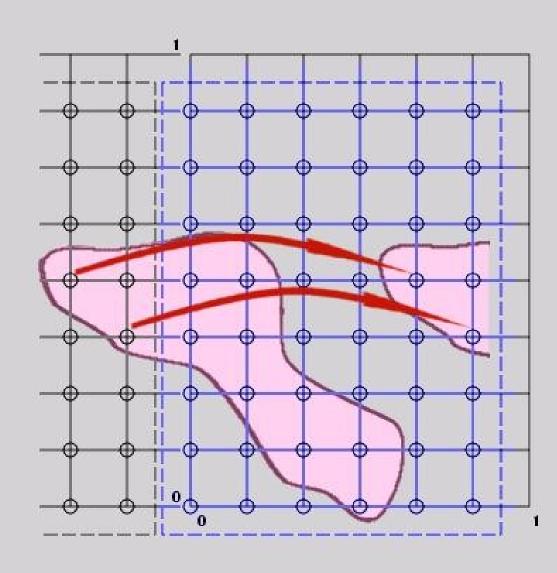


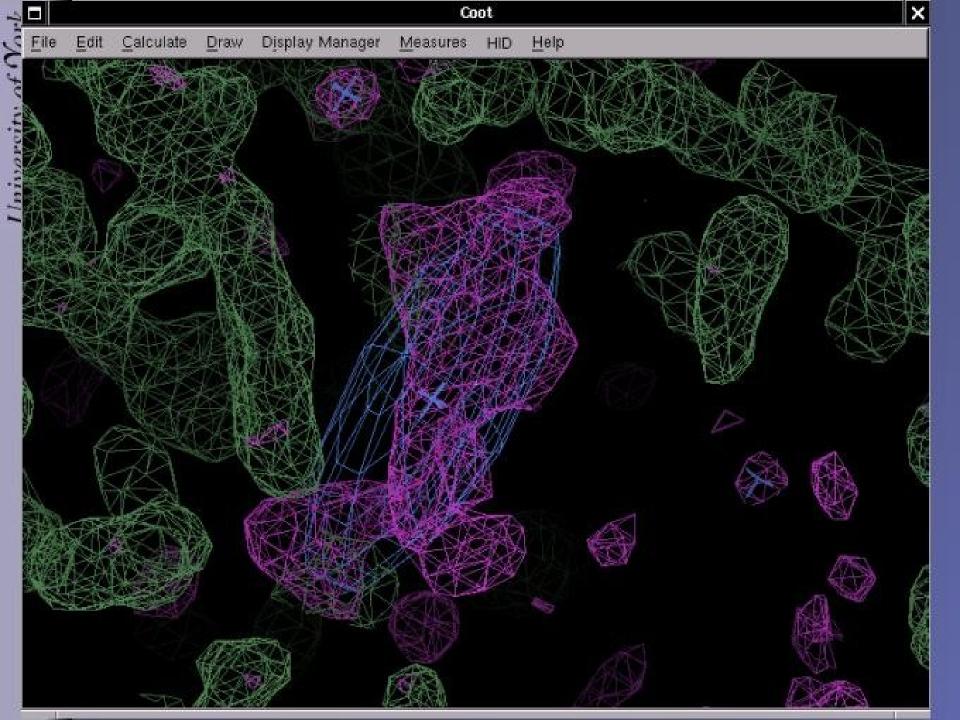
Clipper Map Mapping

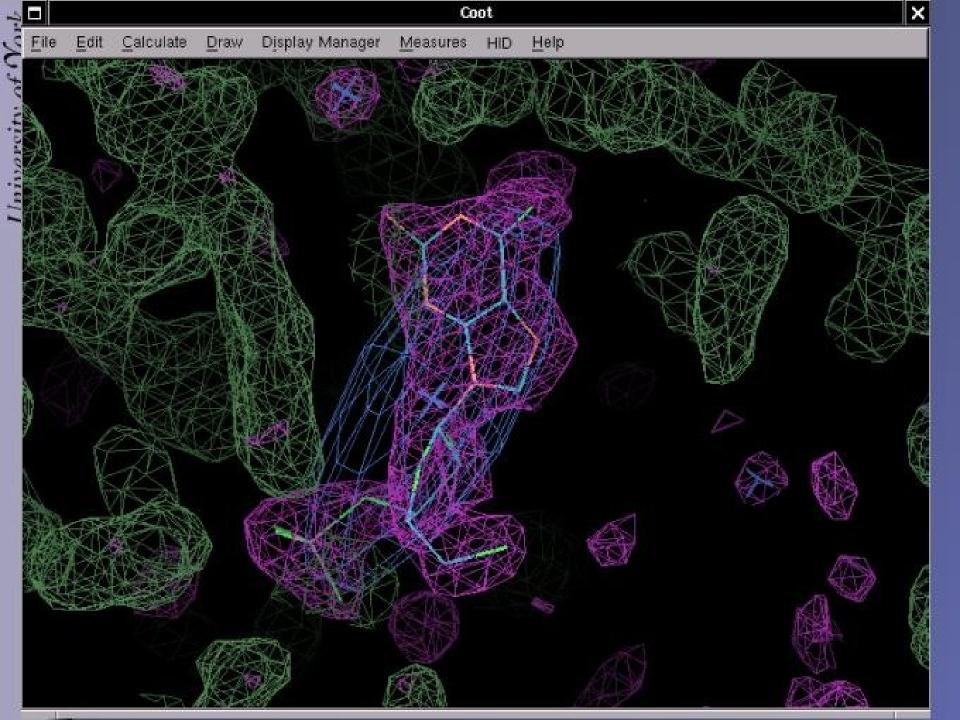
- Clipper maps
 - Appear to be "infinite"
 - Density value can be queried anywhere in space

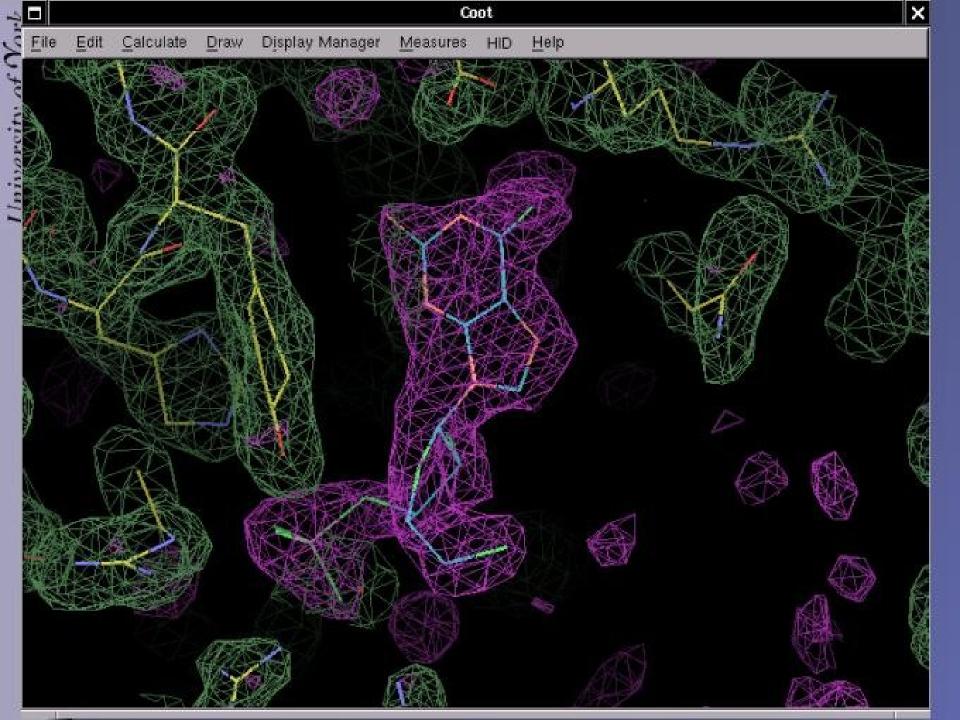












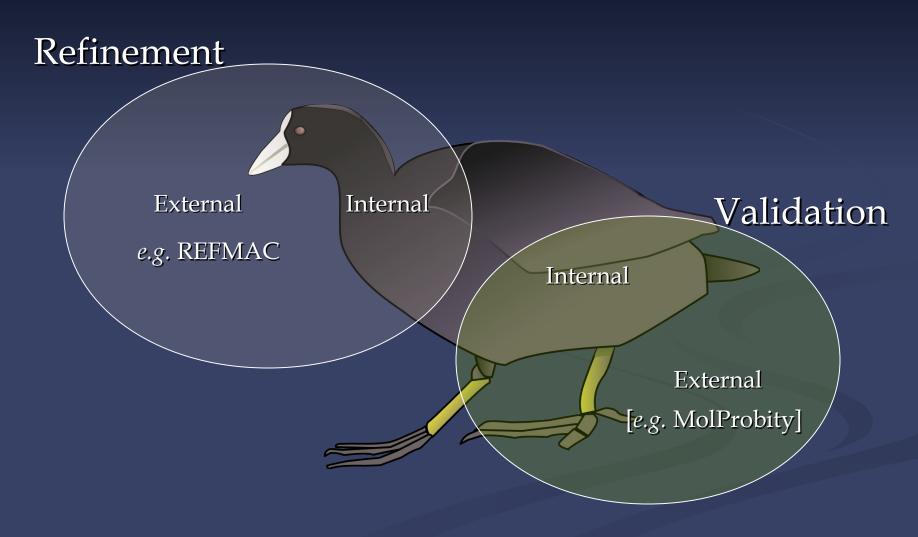
Conformation Idealization

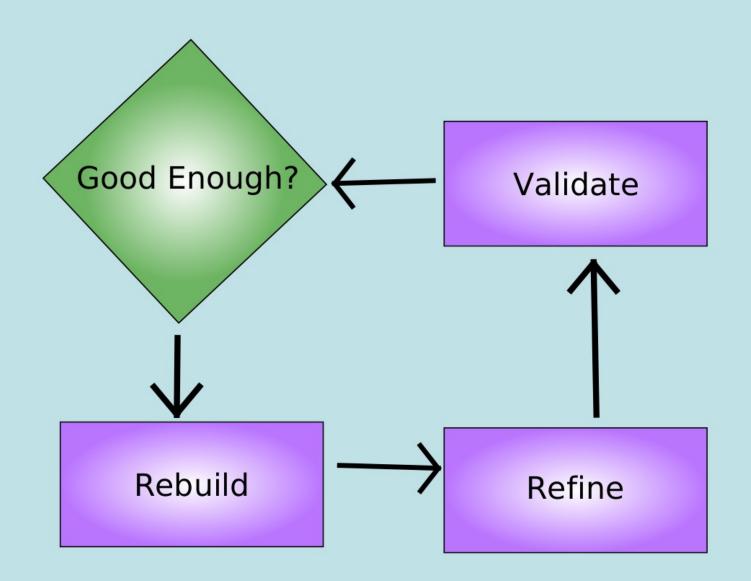
- Each conformer is passed through the "Regularization" function of Coot
 - Non-bonded terms included
- Better to have hydrogen atoms on the model
- Slows things down a good deal...
 - May not be the best method to explore conformational variability for many rotatable bonds

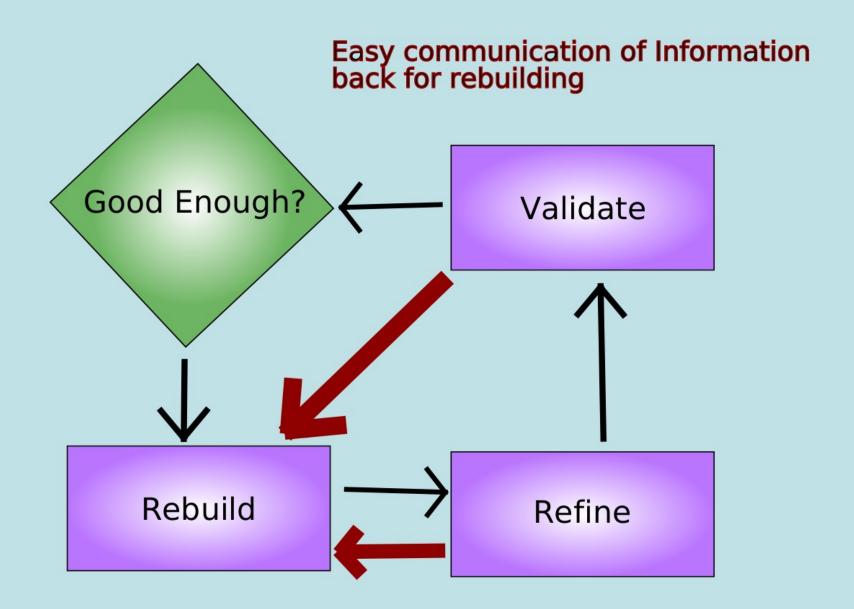
Ligand Overlay

- Algorithm and Code by Eugene Krissinel
- Tries to overlay different ligands/monomers by graph matching
- Useful for "database" ligands where atom names are not selected by hand
- Has been used as the basis of the function which "mutates" residues to alternative monomer types
 - e.g. phosphorylation

Feature Integration







Validation...

What is Validation?

- Comparison of Various aspects of the model with pre-conceived notions of "good quality"
 - Includes unrestrained and restrained criteria
 - Many aspects of validation overlap with refinement and model-building

Why Validate?

- Model-building is error-prone
 - (although automated methods seem to do better)
- Someone else did the model-building
- The model was built several years ago
 - and the notion of "good quality" has changed
- Deposition requires validation

Observations to Parameters Ratio

- Some typical numbers
 - to 2Å, 22000 reflections
 - 200 residues x 10 (atoms/residue) x 4 params/ atom
 - -> about 2.6
- To 3Å:
 - Ratio is about 1:1
- As statisticians, we prefer our models to be parsimonious

Depending on solvent content and the manner in which NCS is handled

A "good" model

- Makes statistical sense
 - The reciprocal space representation agrees tolerably well with the observations (R-factor)
 - No meaningful difference map peaks
- Makes Chemical sense
 - Model Geometry is consistent with the restraints
 - Ramachandran Plot has less than 1% outliers
 - A good clashscore
- Makes Biological sense
 - Residues in chemically sensible environment
 - Is consistent (on the whole) with external biochemistry observations (active site residues)

Quick Bayes

- Bayes Eq:
- Pr(model | data) ∝ Pr(data | model) * Pr(model)
- Pr(data | model) is also called the Likelihood, L(model | data)

Validation Tools - Pr(model)

- Ramachandran Plot
 - Kleywegt Plot (NCS differences)
- Geometry Analysis
- Peptide ω Analysis
- Temperature Factor Analysis
- Rotamer Analysis
- [Clashes]

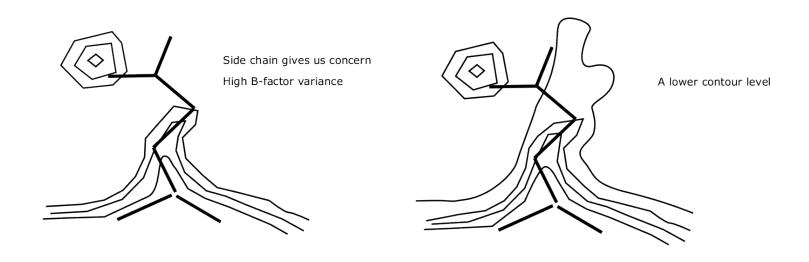
Rotamers

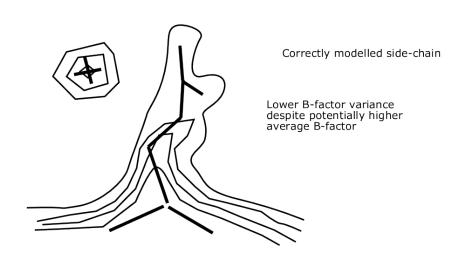
- Side-chains have certain preferred combinations of torsions round their rotatable bonds
- An analysis (batched around the staggered conformations) will give rotamer occurrence

Validation Tools - Pr(data|model)

- Density Fit Analysis
- Difference Map Peaks
 - Variance analysis at Water Positions
- Unmodelled blobs

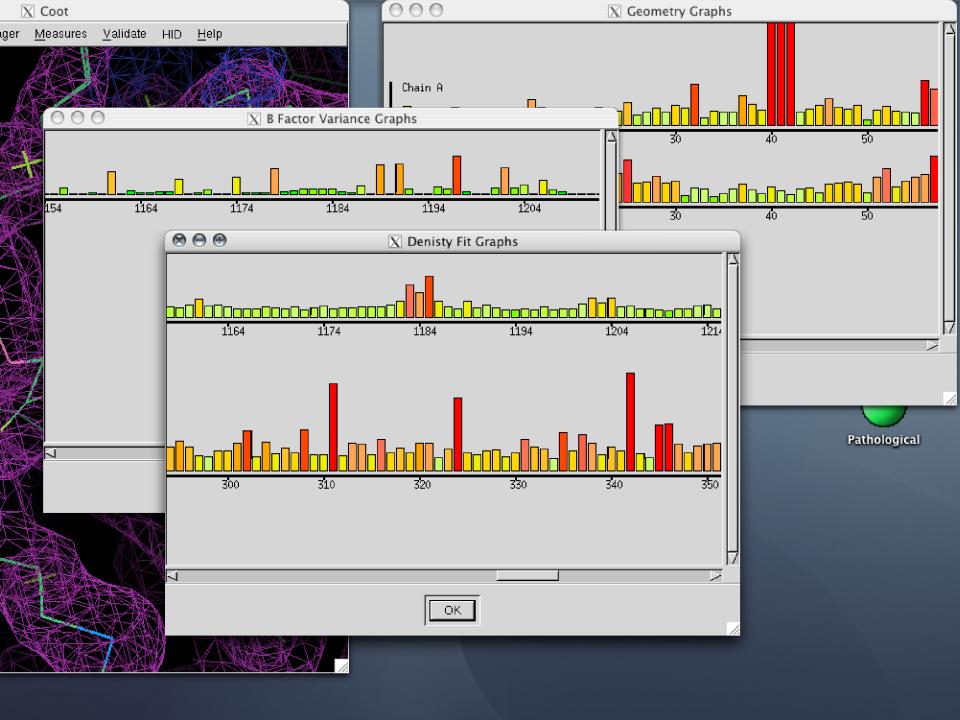
B-factor variance





Chiral Volume Analysis

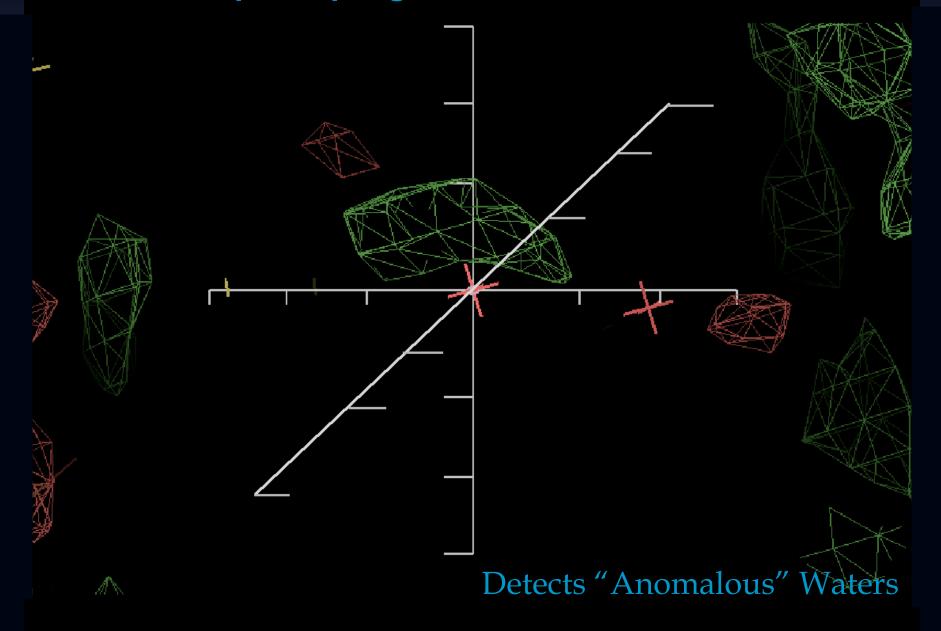
- Based on data in the Refmac dictionary
- ...was needed because it was possible with Coot to accidentally invert Chiral centres
 - e.g. Cas, C β (THR)
 - (Easily corrected with the Mutate & Autofit tool)
 - These days we have chiral volume restraints

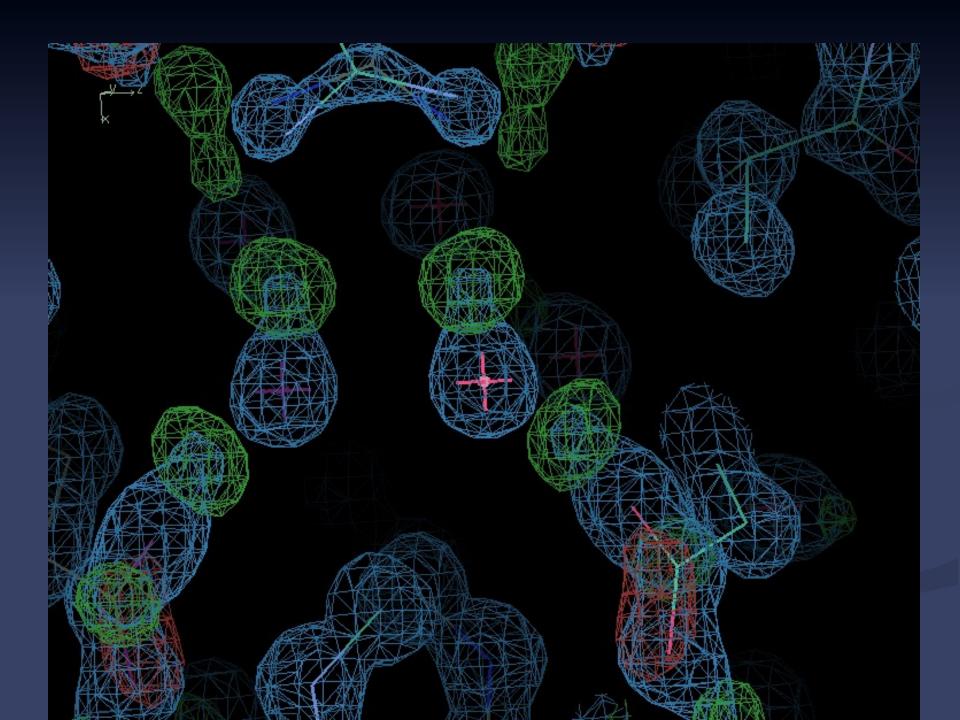


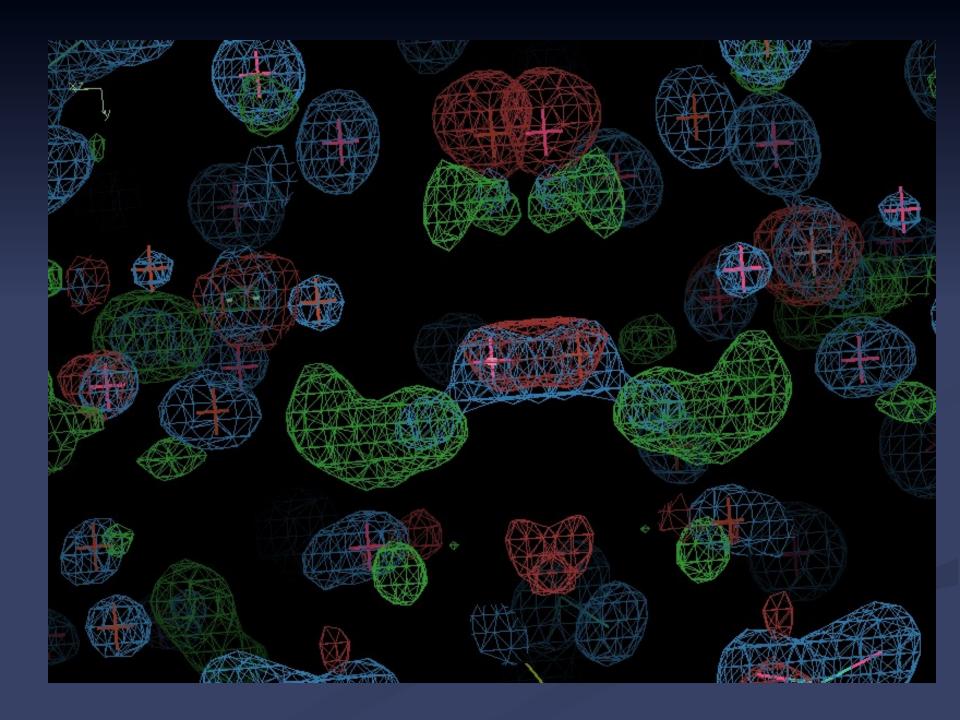
Check/Delete Waters

$\Theta \Theta \Theta$	X Chec	king Waters	:			
Molecule Number: 2aulemsley/data/pad/pad15_8.pdb						
Action: Ger	nerate a List 🔟					
with B factor greater than: 80.00 A^2				☐ Active		
with map sigma level less than: 1.00 electrons/A3				☐ Active		
with closest contact less than: 2.30 A				☐ Active		
with closest	☐ Active					
☐ Ignore Partial Occupancy close contacts						
☐ Ignore Waters with Zero Occupancy						
For Waters that match:						
♦ Any Criterion						
→ All Criteria						
	ок		Cancel			

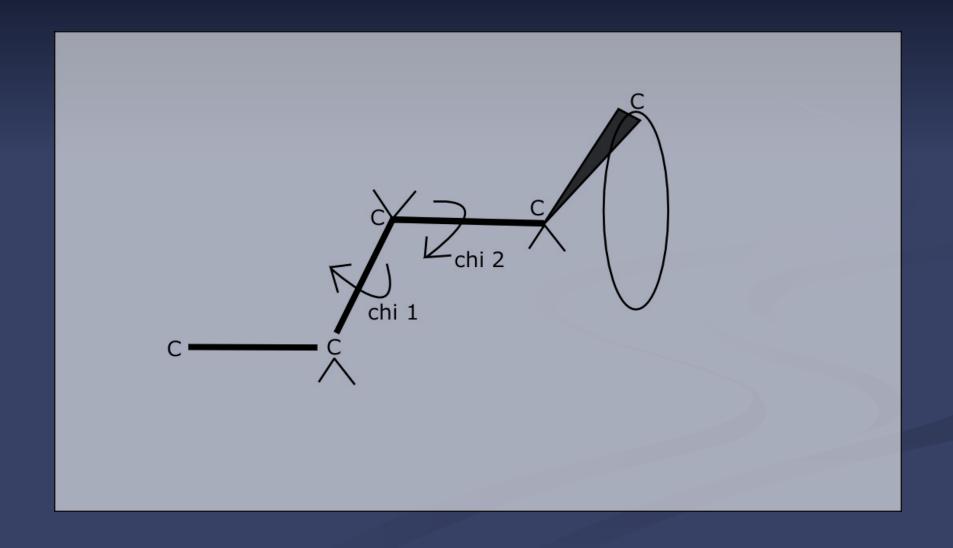
Difference Map Sampling

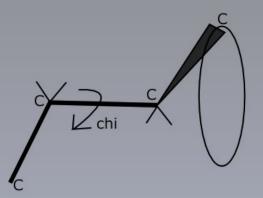






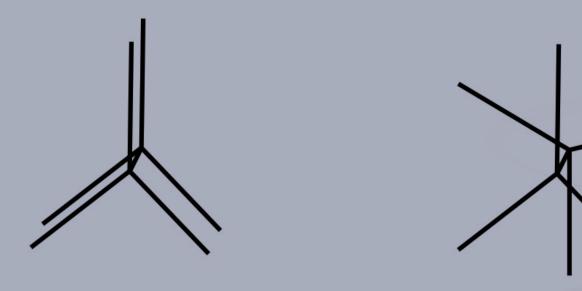
Torsion-based Validation





In principle, there is free rotation

In practice, staggered is energetically more favoured

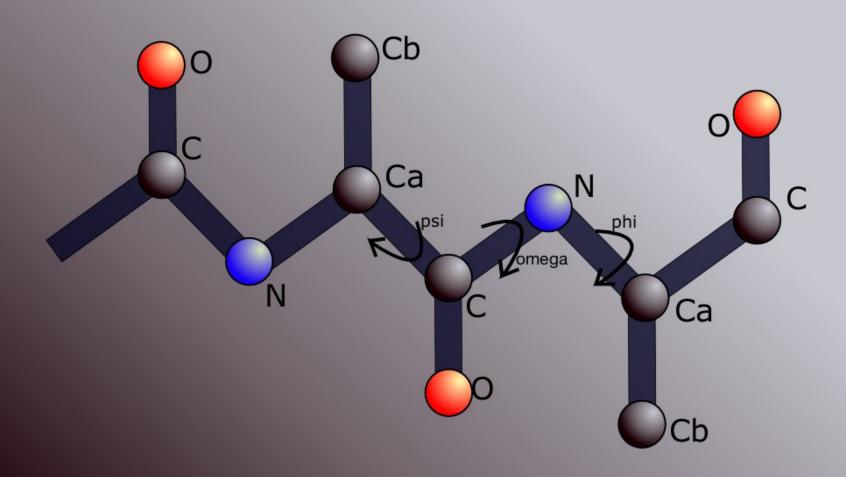


Eclipsed

Staggered

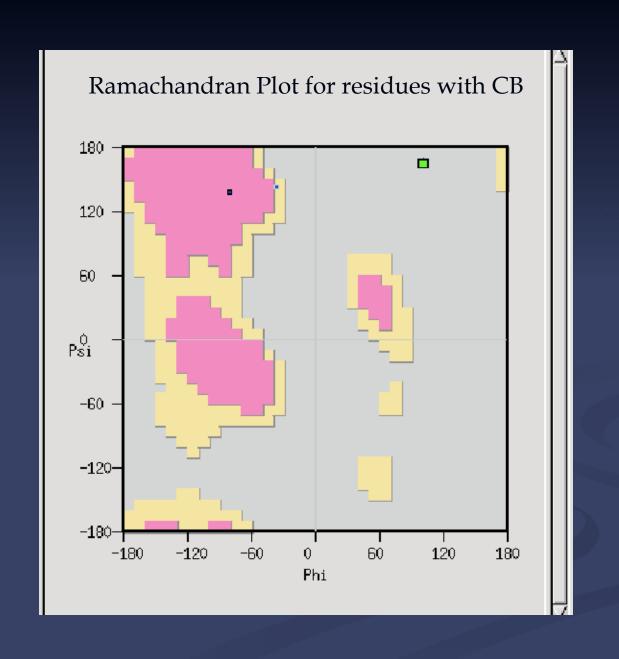
Most favoured staggering angles 60, 180, -60 degrees

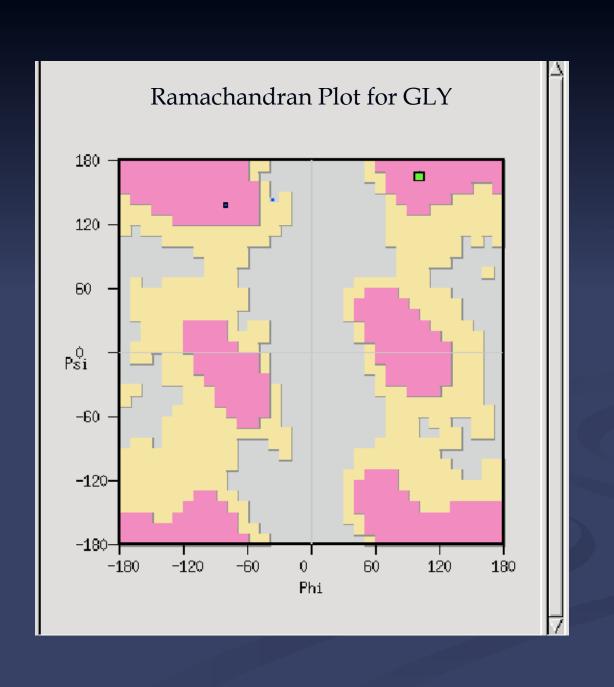
Peptide Torsion Angles

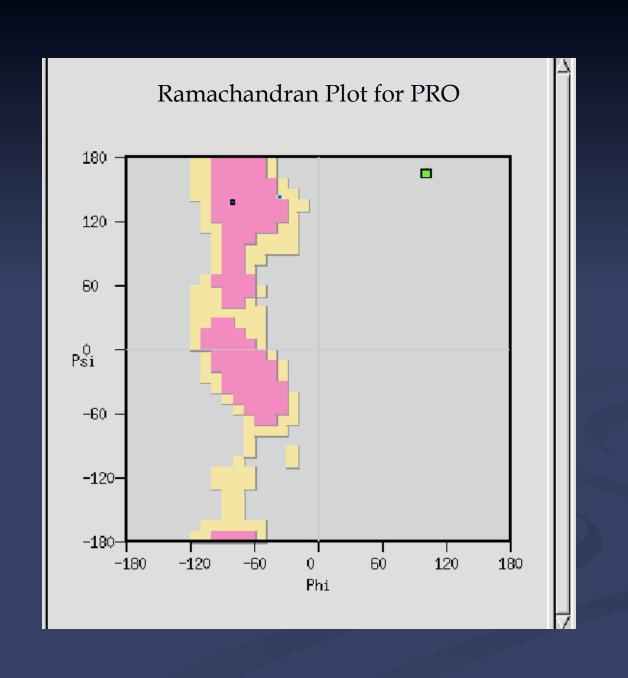


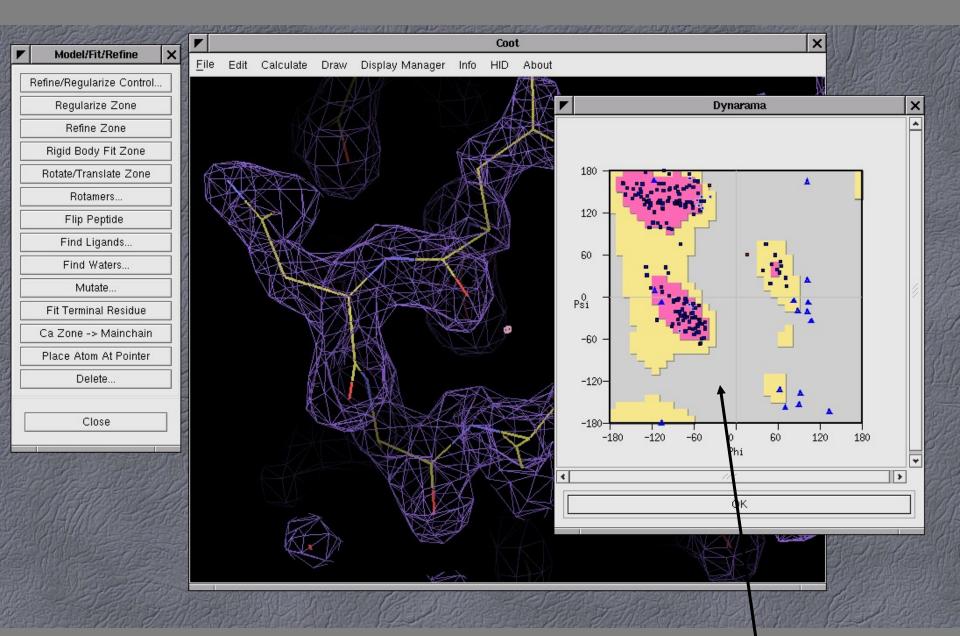
Peptide w

- Needed to check the planarity of the peptide link
 - At low resolutions it is possible to give the protein lots of (too much) freedom to optimize the fit to the density
- Can accidentally create CIS peptides
 - When discovered they are easily reconverted using the CIS<->TRANS peptide tool
- Less accidents happen when peptide plane restraints are applied



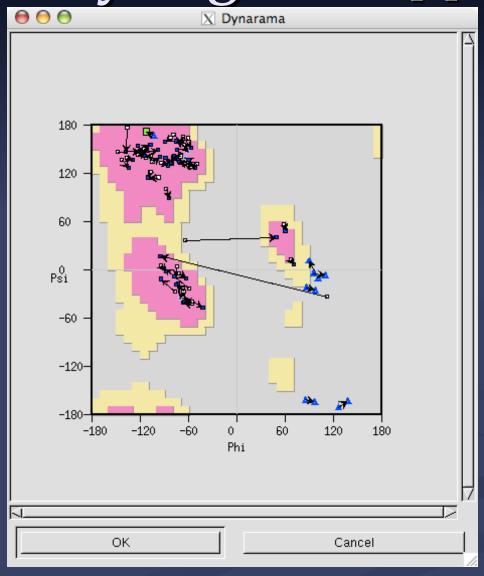






Top500-based distribution

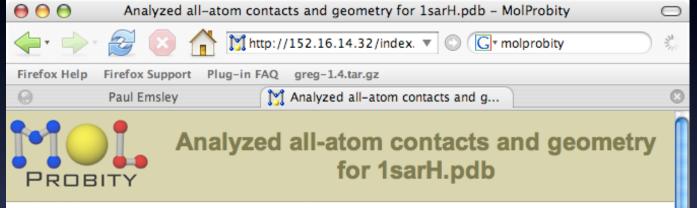
Kleywegt Plots[*]



[*] Named by George Sheldrick

More Validation Pr(model)

- Coot has interface to Molprobity
 - (Molprobity is the widely regarded as the best model validation suite)
- Uses identical Ramachandran plot
- Uses identical Rotamer library
- Coot reads probe dots directly



Analyzed all-atom contacts and geometry for 1sarH.pdb

Entry begun: Today at 1:49pm EST Last modified: Today at 1:49pm EST

Summary statistics

All-Aloili	Clashscore, all atoms:	12.49	60 th percentile* (N=837, 1.55Å - 2.05Å)
	Clashscore, B<40:	10.76	41st percentile* (N=837, 1.55Å - 2.05Å)
Protein Geometry	Rotamer outliers	1.83%	Goal: <1%
	Ramachandran outliers	0.00%	Goal: <0.2%
	Ramachandran favored	99.47%	Goal: >98%
	Cβ deviations >0.25Å	11	Goal: 0
	MER [ALPHA TEST - don't ask]	1.81	78 th percentile* (N=11444, 1.55Å - 2.05Å)

 ^{100&}lt;sup>th</sup> percentile is the best among structures of comparable resolution; 0th percentile is the worst.

Multi-criterion visualizations

Multi-criterion kinemage (970 Kb): View in KiNG | Download

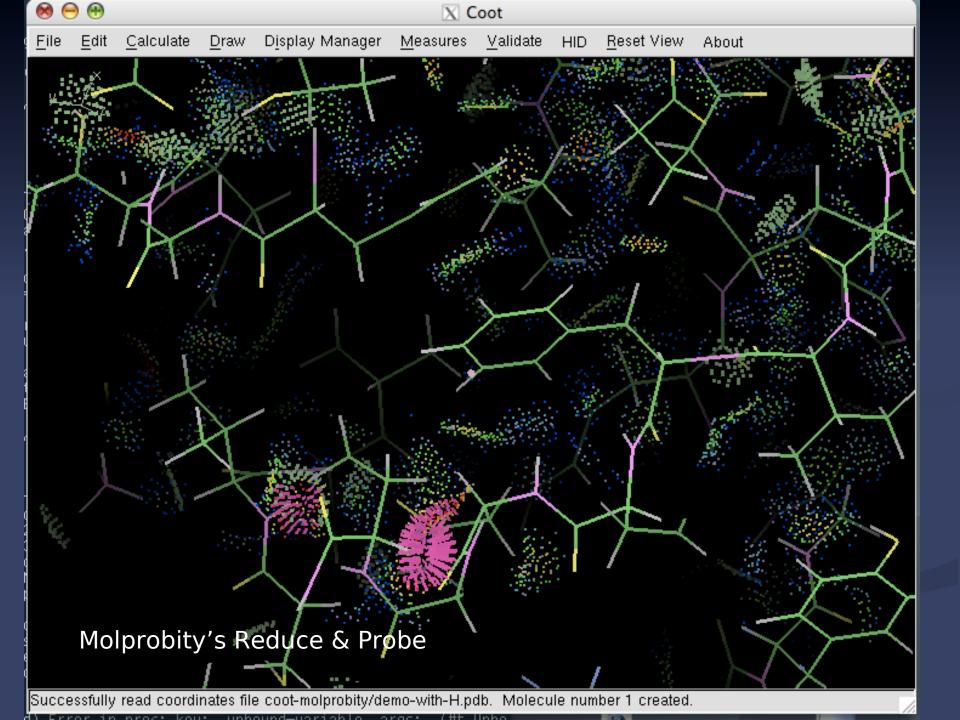
Multi-adiamon char

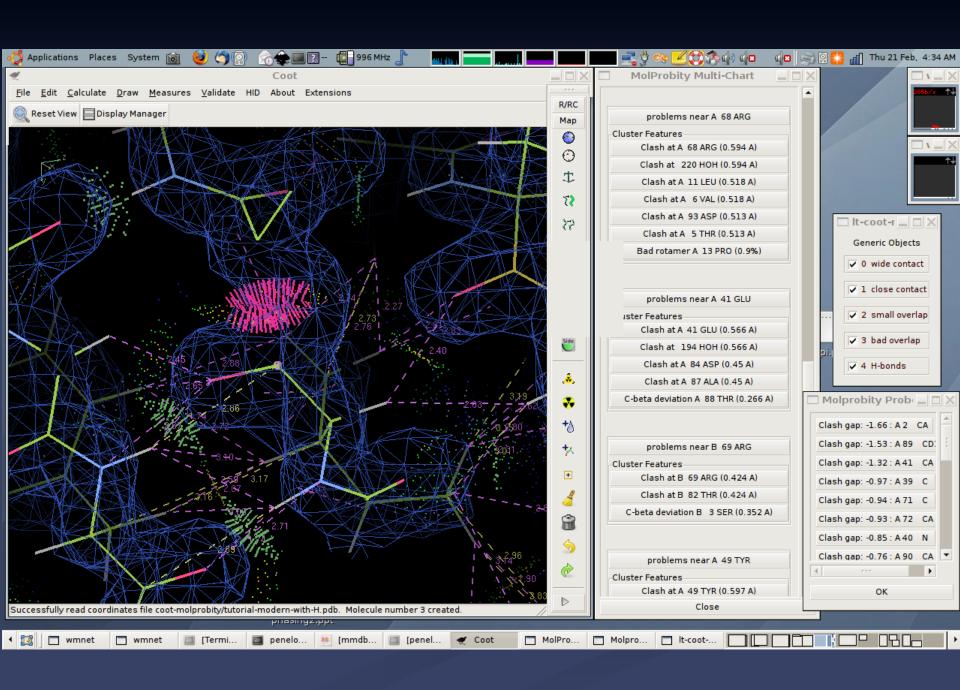
Download multi-criterion to-do list for Coot [ALPHA TEST] Open this in Coot using Calculate | Run Script...

Single-criterion visualizations

- Clash list
- Ramachandran plot kinemage (344 Kb): View in KiNG | Download
- Ramachandran plot PDF
- Cβ deviation scatter plot (2D) (19 Kb): View in KiNG | Download







Other Programs

- Moprobity Suite
 - molprobity.biochem.duke.edu
- WHATCHECK
- VERIFY-3D

Acknowledgements

- Paul Emsley
- Kevin Cowtan
- Eleanor Dodson
- Keith Wilson

http://www.ysbl.york.ac.uk/~emsley/coot

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Google: Coot

or for WinCoot

http://www.ysbl.ac.uk/~lohkamp/coot

- Libraries, dictionaries
 - Alexei Vagin, Eugene Krissinel, Stuart McNicholas
 - Dunbrack, Richardsons
- Coot Builders and Testers
 - William Scott, Ezra Peisach
 - York YSBL, Dundee, Glasgow (early adopters)
 - Coot Mailing List subscribers