

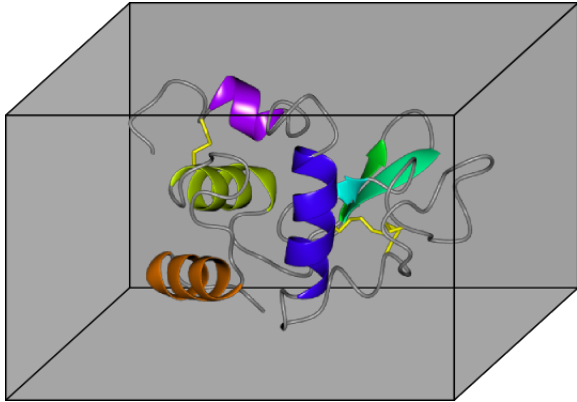
Molecular Replacement

Andrey Lebedev

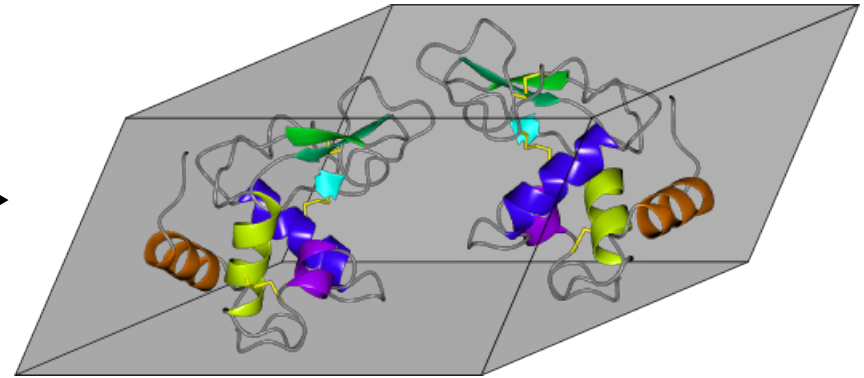
CCP4

MR Problem

Known crystal structure



New crystal structure



Given:

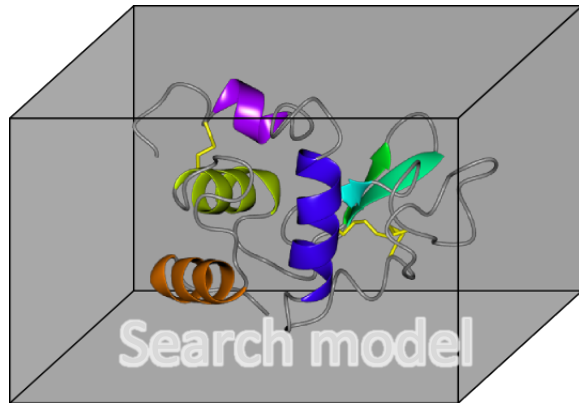
- Crystal structure of a homologue
- New X-ray data

Find:

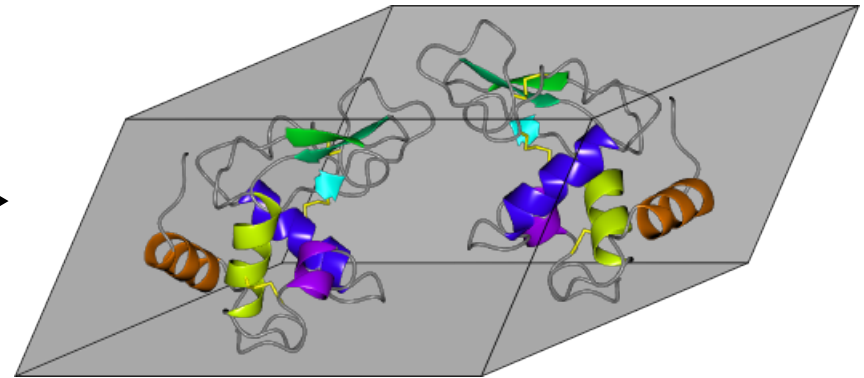
- The new crystal structure

MR Technique

Known crystal structure



New crystal structure



Method:

- $6 \times N$ - dimensional global optimisation
 - one 6-d search for each molecule in the AU
 - >> split further to orientation + translation searches = 3 + 3
 - >> fast search step using FFT

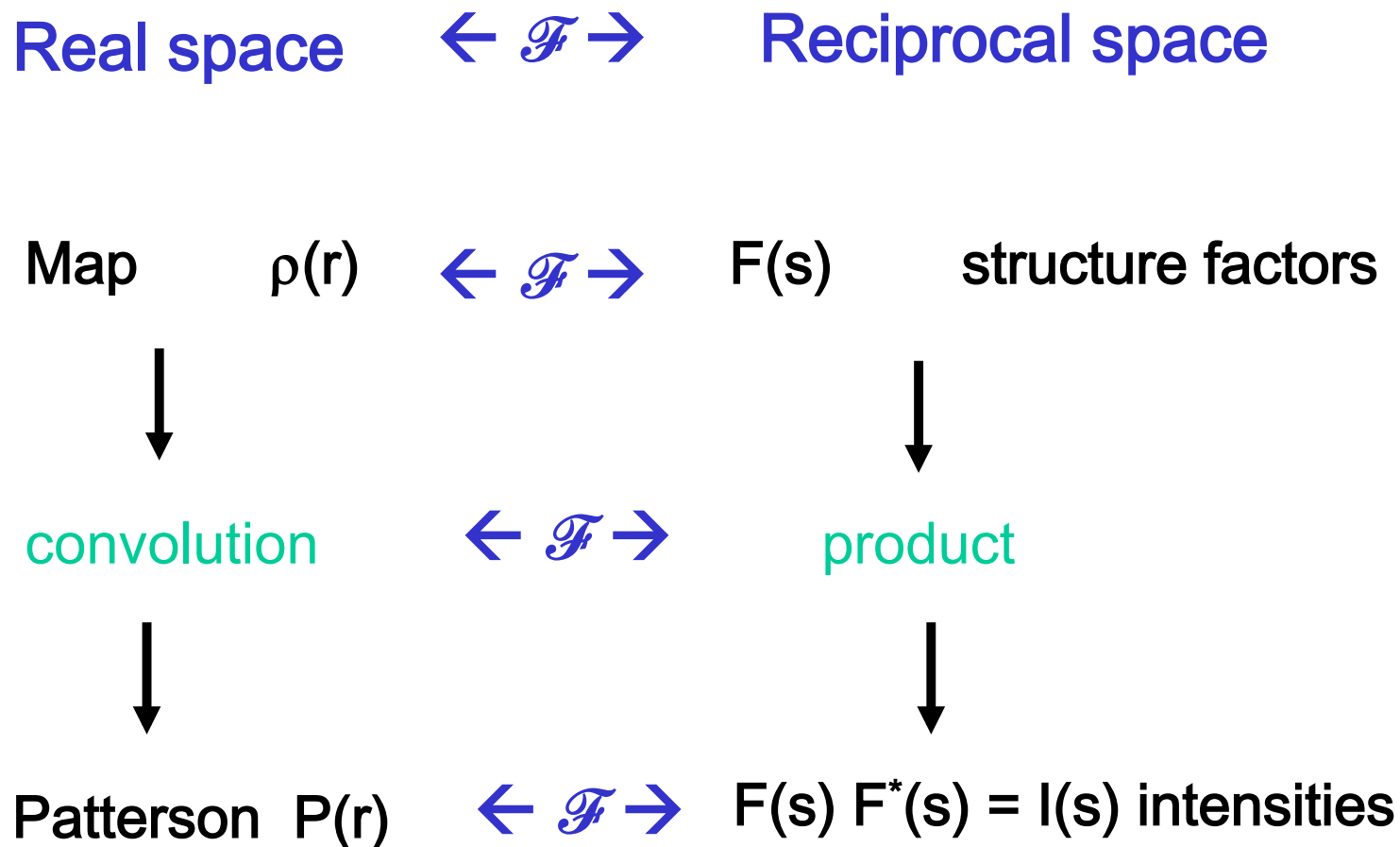
Required:

- Scoring
 - the match between the data and an (incomplete) crystal model
 - ideally: the highest score = correct solution

Real and Reciprocal spaces

- Terms may refer real space but actual calculations may be performed in the reciprocal space:
 - "Search in the electron density"
 - "Patterson search"
- The concepts formulated in real space are more intuitive

Functions in Real and Reciprocal spaces



Structure factors and Electron density map

Structure factors $F(h,k,l)$

- A discrete complex function in the reciprocal space

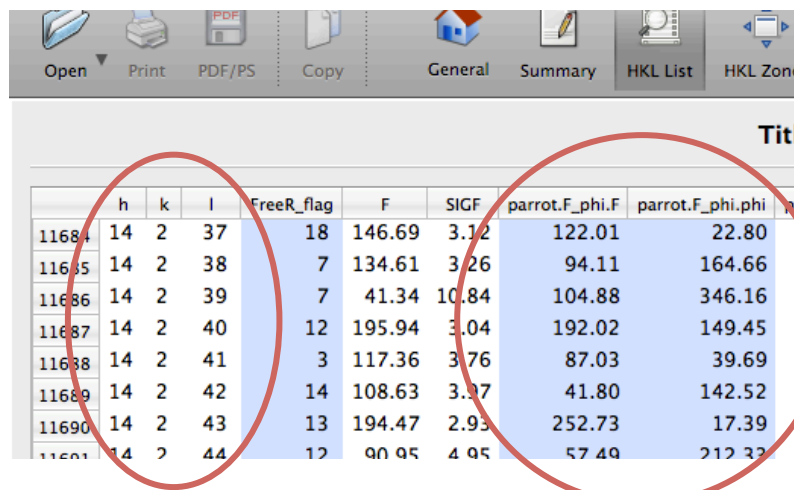
At given h, k, l

- Complex number:

$$F = A + iB$$

- Can be expressed via structure amplitude and phase

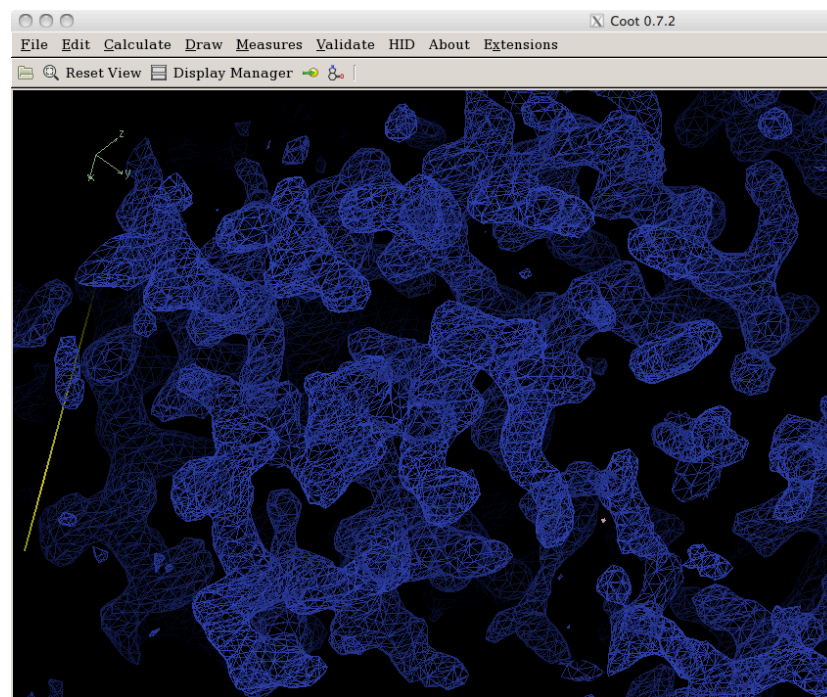
$$F = |F| \exp(i\phi)$$



	h	k	l	FreeR_flag	F	SIGF	parrot.F_phi.F	parrot.F_phi.phi
11684	14	2	37	18	146.69	3.12	122.01	22.80
11685	14	2	38	7	134.61	3.26	94.11	164.66
11686	14	2	39	7	41.34	10.84	104.88	346.16
11687	14	2	40	12	195.94	3.04	192.02	149.45
11688	14	2	41	3	117.36	3.76	87.03	39.69
11689	14	2	42	14	108.63	3.07	41.80	142.52
11690	14	2	43	13	194.47	2.93	252.73	17.39
11691	14	2	44	12	90.95	4.95	57.49	212.33

Electron density map

- periodic 3-d function in real space



is directly interpretable

- model building
- real-space fitting of fragments

Intensities and Patterson map

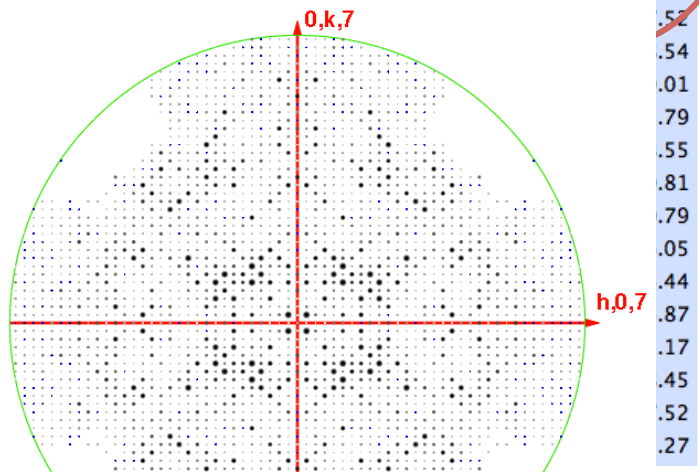
Intensities $I(h,k,l)$

- 3-d discrete real function in the reciprocal space

Open Print PDF/PS Copy General Summary HKL

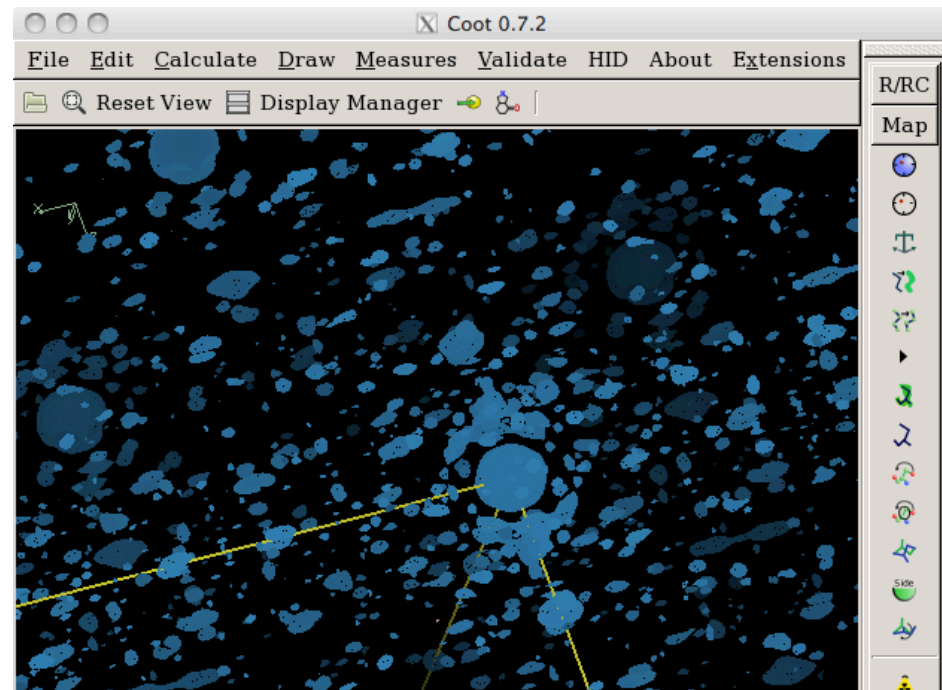
Title: P222

	h	k	l	FREE	F	SIGF	IMEAN	SIGMEAN
13671	12	9	34	15	145.37	38.61	54.57	45.85
13672	12	9	35	11	144.03	38.25	54.00	45.05
13673	12	10	0	15	131.82	49.54	68.47	44.37
13674	12	10	1	13	88.30	28.53	-38.02	29.04
13675	12	10	2	13	239.79	29.58	156.79	33.71
13676	12	10	3	3	356.73	21.77	328.07	38.83
13677	12	10	4	3	112.79	33.04	8.13	18.69



Patterson map:

- 3-d function in real^(*) space



- Nothing reminds a protein molecule
- **Model building**, residue by residue, is **impossible**

MR: Two distinct cases dependent on availability of phases

- Data = structure factors (**include phases**)
 - "**Search in the electron density**"
 - Electron density maps are compared: calculated vs. observed
 - Model building is a more straightforward approach
 - » Useful in special cases
- Data = observed intensities (**no phases**)
 - "**Patterson search**"
 - Patterson maps are compared: calculated vs. observed
 - Direct model building is impossible in the absence of phases
 - » The most common case of MR

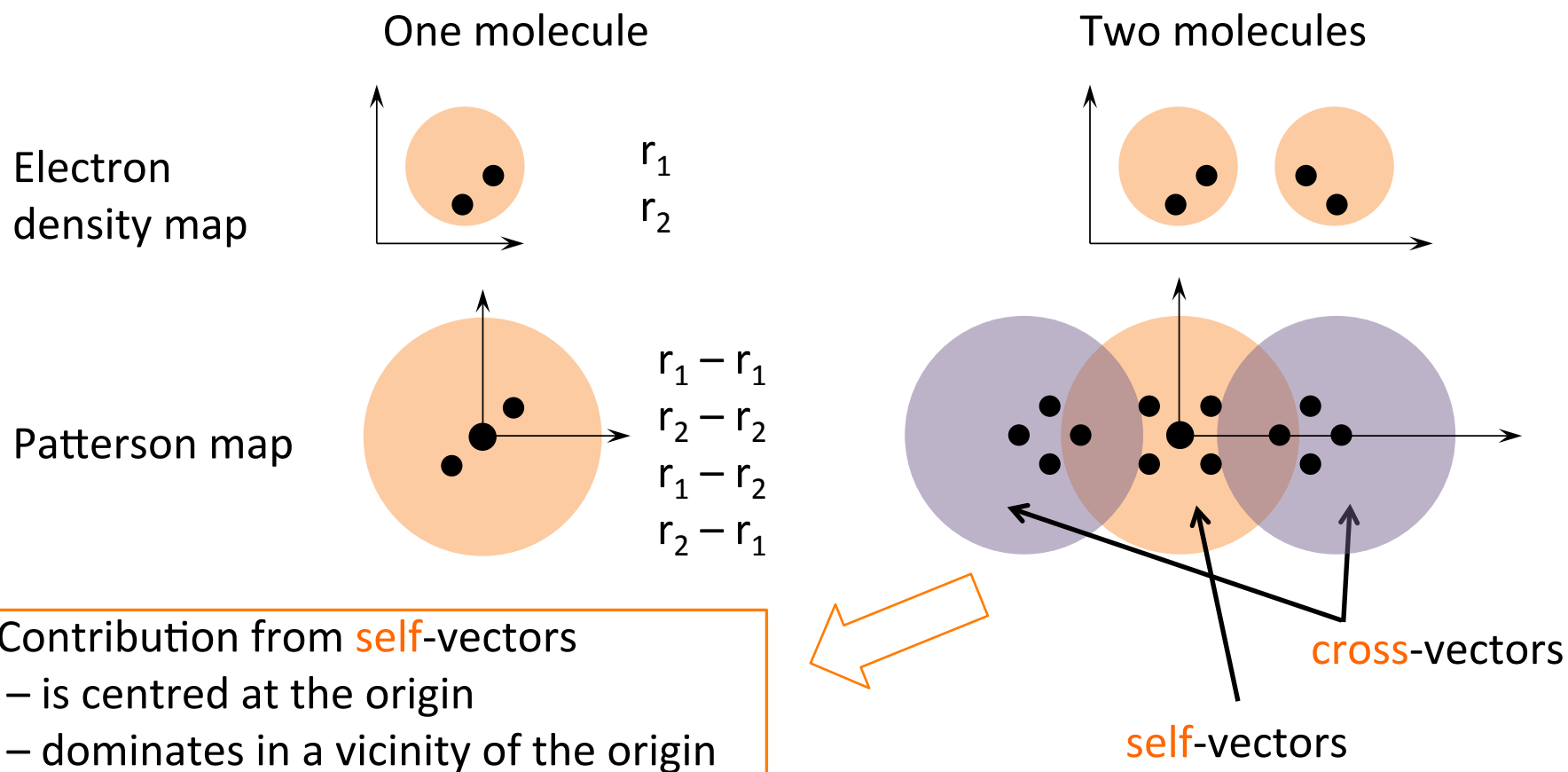
As a rule, all computations are in the reciprocal space

Self and cross vectors

Electron density map = peaks from all atoms

Patterson map = peaks from all interatomic vectors

- **self**-vectors: vectors between atoms belonging to the same molecule
- **cross**-vectors: vectors between atoms belonging to different molecules



What can be seen to be separated?

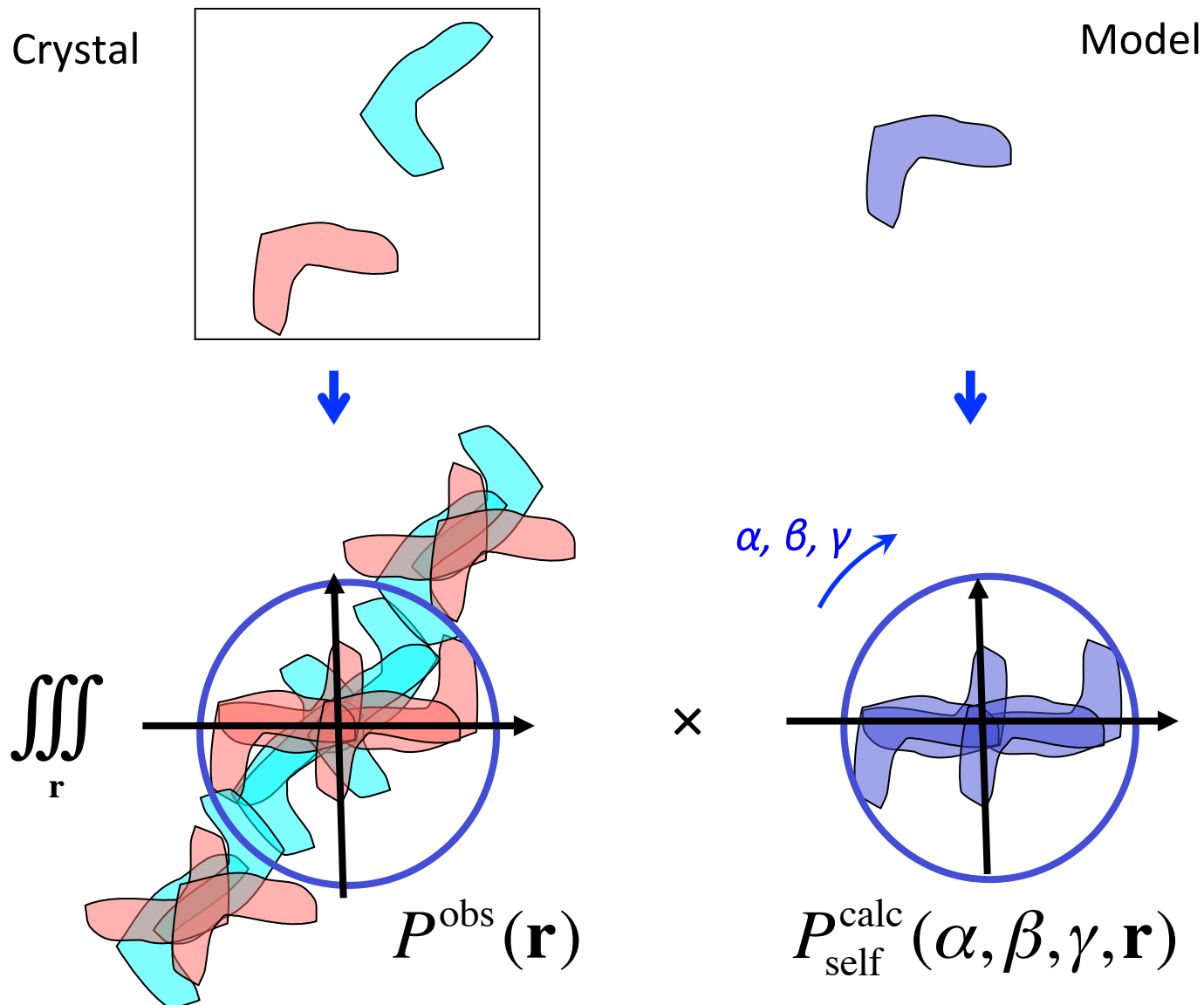
Electron density maps:

- Peaks from atoms or larger fragments are separated in space
 - Model building is possible

Patterson map:

- Contribution from **self**-vectors is centred at the origin
- **Self**-vectors are, in average, shorter than **cross**-vectors
 - Peaks from **self**-vectors dominates in a vicinity of **the origin**
 - Peaks from **cross**-vector dominates **away from the origin**
- One 6-dimensional search splits into
 - Rotation Function: 3-dimensional search (using **self**-vectors)
 - Translation Function: 3-dimensional search (using **cross**-vectors)

Rotation Function



Rotation Function

$$RF(\alpha, \beta, \gamma) = \iiint P^{\text{obs}}(\mathbf{r}) \times P_{\text{self}}^{\text{calc}}(\alpha, \beta, \gamma, \mathbf{r}) d\mathbf{r}^3$$

$P_{\text{self}}^{\text{calc}}(\alpha, \beta, \gamma, \mathbf{r})$ contains only

- self-vectors

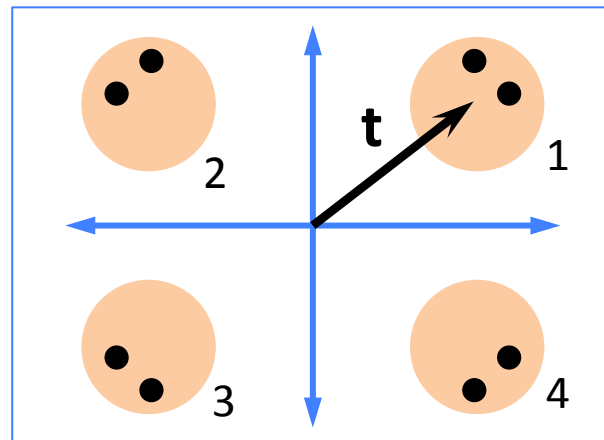
$P^{\text{obs}}(\mathbf{r})$ contains

- self-vectors (signal from one of the orientations in the crystal)
- cross-vectors (noise)

Translation Function

Analytical steps

- The centre of molecule 1
 - Parameter \mathbf{t}
- Centres of molecules 2, 3 and 4
 - form symmetry operations



- $F_{\mathbf{h}}^{\text{calc}}(\mathbf{t})$, $I_{\mathbf{h}}^{\text{calc}}(\mathbf{t})$
- $TF(\mathbf{t}) = \sum_{\mathbf{h}} I_{\mathbf{h}}^{\text{obs}} \times I_{\mathbf{h}}^{\text{calc}}(\mathbf{t})$
- Can be converted to a form:

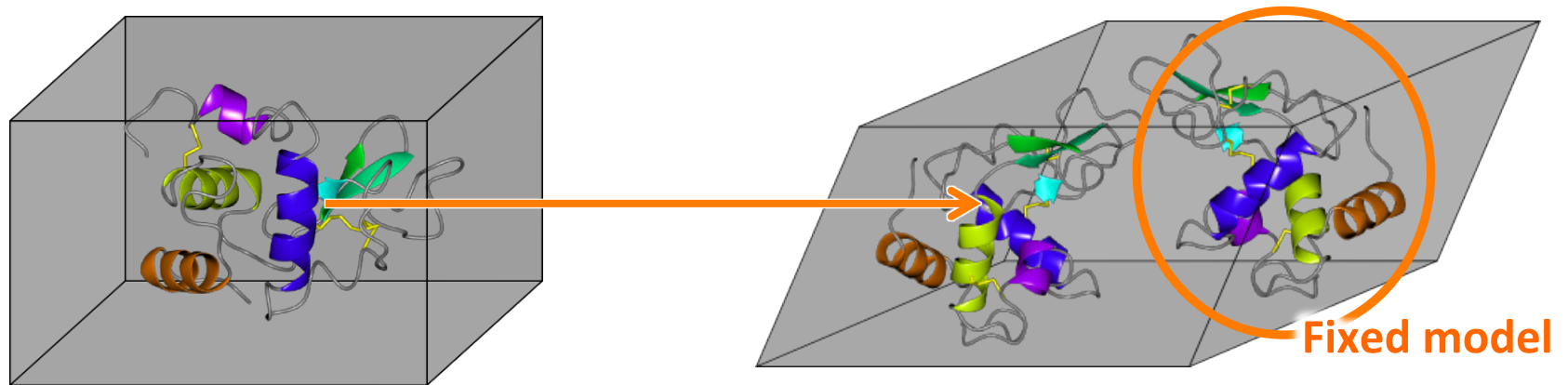
$$TF(\mathbf{t}) = \sum_{\mathbf{h}} G_{\mathbf{h}} \exp(2\pi i \mathbf{h} \cdot \mathbf{t})$$

- FFT techniques can be applied

Numerical calculations

- $G_{\mathbf{h}}$
- FFT: $G_{\mathbf{h}} \rightarrow TF(\mathbf{t})$
- Peak search in $TF(\mathbf{t})$: best \mathbf{t}

Fixed partial model



Almost the same equation as for a single molecule search,

$$TF(\mathbf{t}) = \sum_{\mathbf{h}} I_{\mathbf{h}}^{\text{obs}} \times \left| F_{\mathbf{h}}^{\text{fixed}} + F_{\mathbf{h}}^{\text{calc}}(\mathbf{t}) \right|^2$$

Again, can be converted to a form:

$$TF(\mathbf{t}) = \sum_{\mathbf{h}} G_{\mathbf{h}} \exp(2\pi i \mathbf{h} \cdot \mathbf{t})$$

and **FFT** technique can be used

Translation Function

$$TF(\mathbf{t}) = \iiint P^{\text{obs}}(\mathbf{r}) \times P_{\text{cross}}^{\text{calc}}(\mathbf{t}, \mathbf{r}) d\mathbf{r}^3$$

$P_{\text{cross}}^{\text{calc}}(\mathbf{t}, \mathbf{r})$ contains

- **cross**-vectors

$P^{\text{obs}}(\mathbf{r})$ contains

- **self**-vectors (background or **noise**)
- **cross**-vectors (relevant vectors: **signal**, others: **noise**)

Packing considerations

Molecules in the crystal **do not overlap**

How can we use this information?

» Patterson map does not explicitly reveal molecular packing

Reject MR solutions

- Restrict distance between centres of molecules
- Count close interatomic contacts

Modify TF

- Divide by Overlap Function
- Multiply by Packing function

Fast Packing Function

Estimation of overlap

- Using mask from search model
- FFT

Packing Function

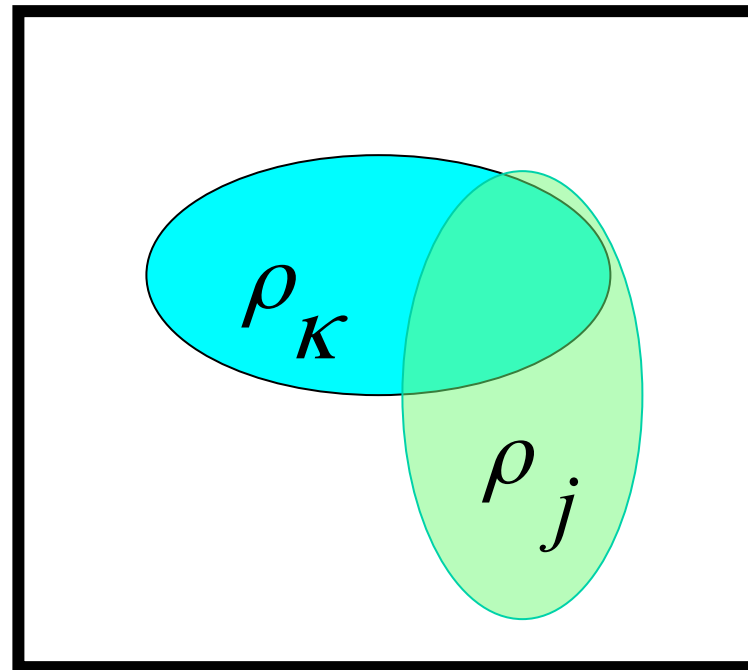
$$PF = 1 - \text{overlap}$$

Modified Translation Function

$$TF_{mod} = TF \times PF$$

Peak search

- Using TF_{mod}
- No irrelevant peaks are passed to rescoring step

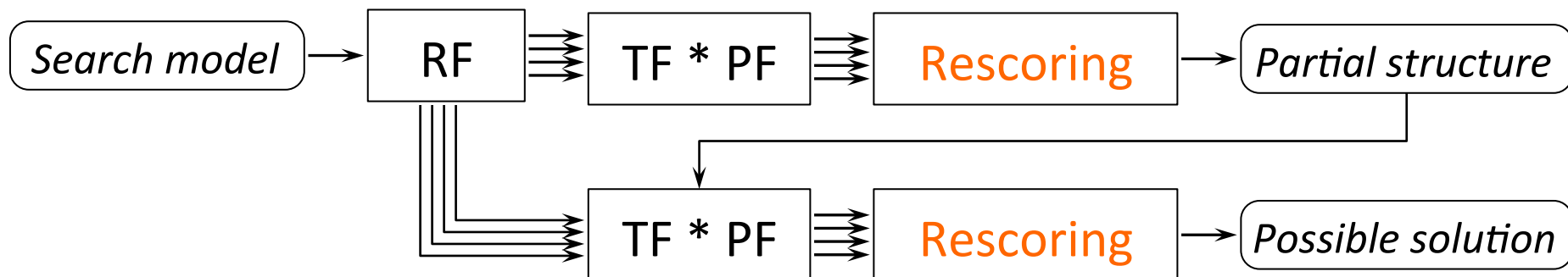


Implemented in *MOLREP*

A conservative MR protocols for two copies of a model

As implemented in *MOLREP*

Intensities \longrightarrow all steps



$$RF = \sum_{hkl} w * I_o * I_c(\alpha\beta\gamma)$$

$$TF = \sum_{hkl} w * I_o * I_c(xyz)$$

Rescoring: Correlation Coefficient* PF

Specialised MR techniques

- Search in the density (phased MR)
- Handling Translational Non-Crystallographic symmetry
 - Non-origin peaks in the Patterson map indicate the presence of TNCS
 - Requires special handling of model errors (Phaser)
 - Molecules related by TNCS can be found in one go as they have nearly the same orientation
- Self Rotation Function
- Locked RF and TF
 - Using point symmetry of oligomers
- Exhaustive searches
- Stochastic searches

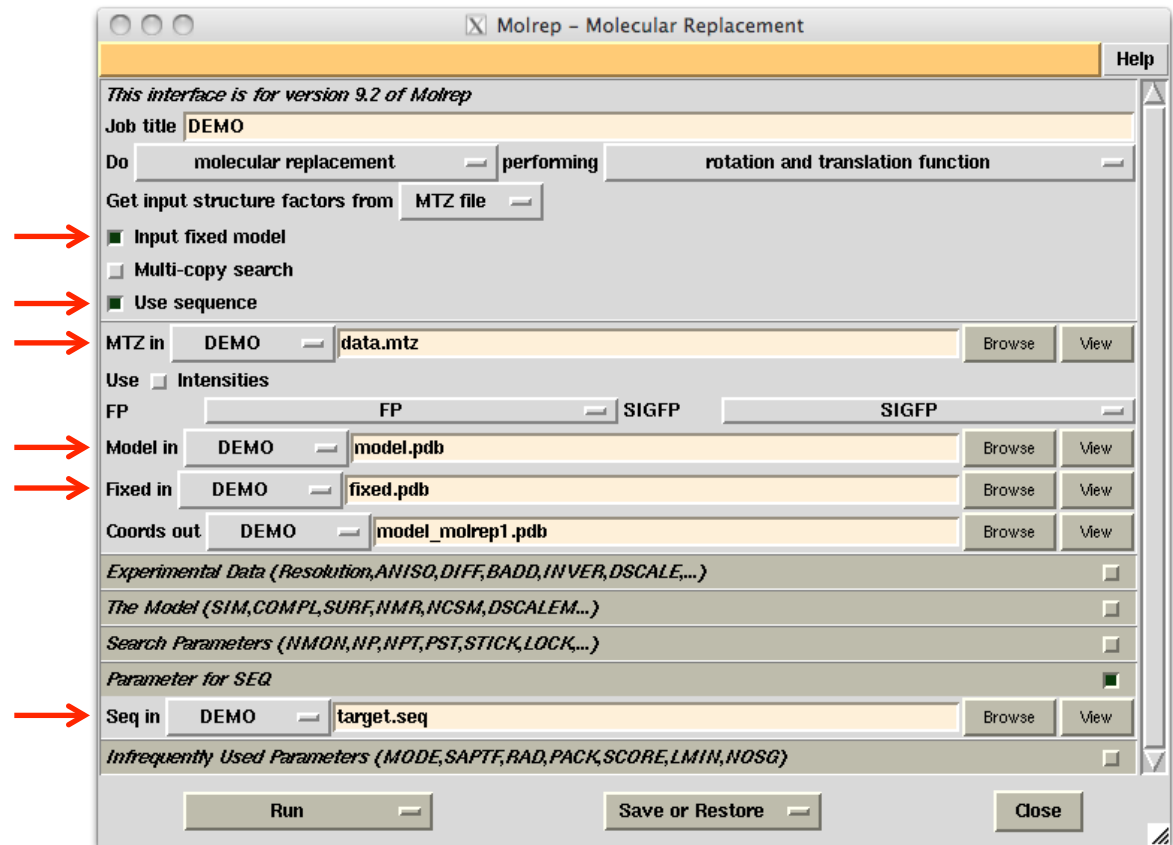
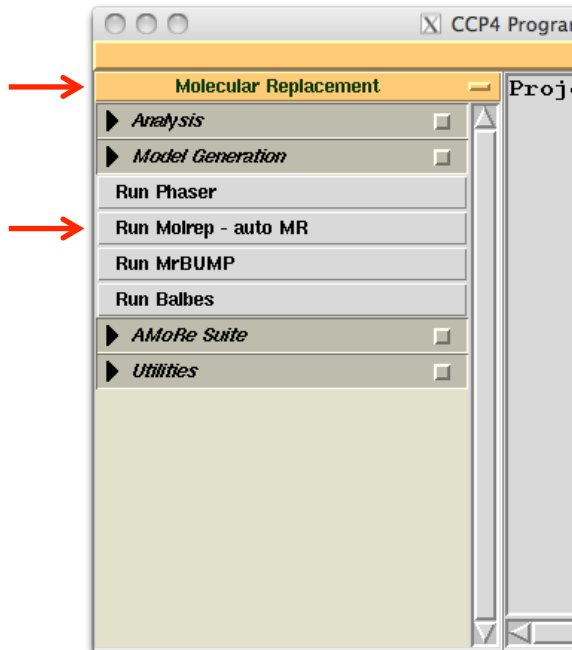
Molrep

Alexey Vagin

YSBL University of York

Molrep

```
molrep -f data.mtz -m model.pdb -mx fixed.pdb -s target.seq
```



Default protocol

```
molrep -f data.mtz -m model.pdb -mx fixed.pdb -s target.seq
```

- model correction if sequence provided
- defines the number of molecules per AU
- modification of the model surface
- anisotropic correction of the data
- weighting the data according to model completeness and similarity
- check for pseudotranslation and use it if present
- 30+ peaks in Cross RF for use in TF (accounts for close peaks)
- applied packing function, $TF \times PF$
- rescoring using $CC \times PF$
- make use of partial structure (fixed model)

Default protocol: log-file

```
CCP4I fileviewer 1_molrep.log
Help

14 4 8.444 2.920 1.00 1.00 -21.06 0.599 0.110 7.33 ( 0.242)
INFO: contrast is good enough. Stop this run

--- Summary ---
+-----+
| RF  TF  theta  phi  chi  tx  ty  tz  TFcnt  wRfac  Score |
+-----+
| 1  1  1  72.59  38.64  179.42  0.825  0.649  0.480  10.06  0.560  0.242 |
| 2  2  1  72.41  38.93  177.39  0.820  0.650  0.480  10.91  0.565  0.217 |
| 3  4  1  72.18  38.99  176.40  0.819  0.652  0.480  9.61  0.573  0.195 |
| 4  7  2  77.85  58.68  142.53  0.445  0.292  0.483  5.03  0.602  0.121 |
| 5  3  2  107.48 -166.00  160.39  0.637  0.790  0.175  4.51  0.599  0.120 |
| 6  6  10  52.26  91.15  50.93  0.416  0.376  0.163  1.95  0.603  0.111 |
| 7  13  12  82.51  133.98  129.34  0.542  0.566  0.253  2.80  0.601  0.110 |
| 8  14  4  81.86  91.66  108.52  0.780  0.260  0.469  2.92  0.599  0.110 |
| 9  9  4  113.57  167.71  124.63  0.757  0.436  0.021  3.39  0.603  0.110 |
| 10 8  13  87.47  114.84  104.62  0.644  0.955  0.369  1.21  0.605  0.109 |
| 11 5  3  108.24 -136.26  176.12  0.816  0.651  0.479  2.79  0.602  0.109 |
| 12 12  1  97.58  104.76  90.32  0.585  0.049  0.166  2.33  0.607  0.107 |
| 13 10  2  98.10  104.76  89.79  0.586  0.049  0.166  1.93  0.607  0.107 |
| 14 11  9  36.40  73.27  110.10  0.394  0.165  0.289  1.16  0.610  0.097 |
+-----+

Contrast = 7.33

After stick correction:
Move closer to origin
I_sym_operator : 11
new position(frac): -0.176 -0.351 0.020

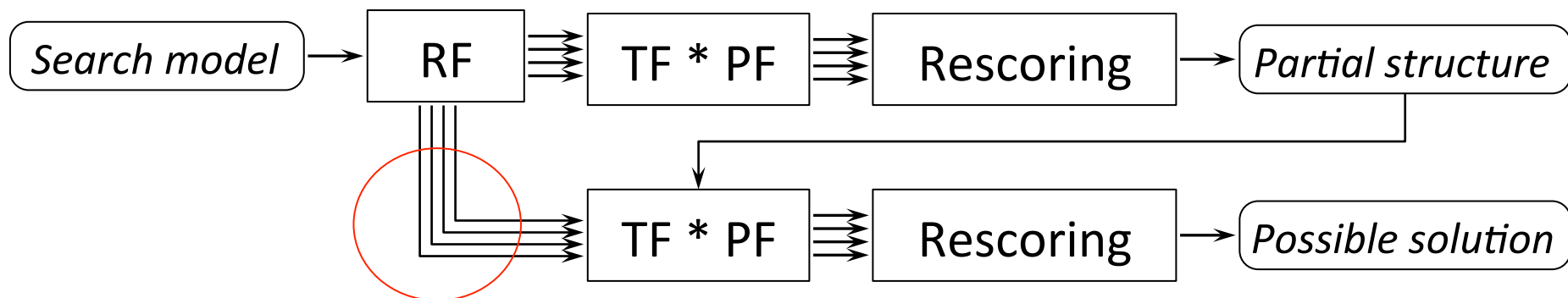
Nmon RF  TF  theta  phi  chi  tx  ty  tz  TFcnt  wRfac  Score
1  1  1  21.87 -179.03  106.86 -0.176 -0.351 0.020  10.06  0.560  0.242

--- convert "molrep.crd" to "molrep.pdb" ---
Time: 1h 27m 58s Elapsed: 0h 0m 57s
MOLREP(ccp4): Normal termination
Times: User: 53.8s System: 2.4s Elapsed: 0.57

Find Show Log Graphs Show Summary Quit
```

Molrep default protocols for two copies of a model

X-ray data \longrightarrow all steps



$$RF = \sum_{hkl} w * I_o * I_c(\alpha\beta\gamma)$$

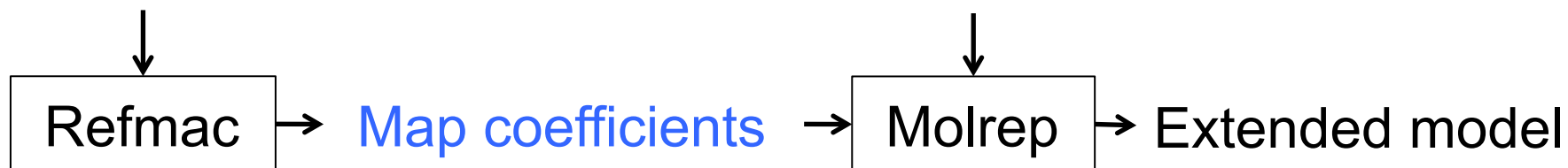
$$TF = \sum_{hkl} w * I_o * I_c(xyz)$$

Rescoring: Correlation Coefficient* PF

Search in the electron density map

Partial structure

Search model



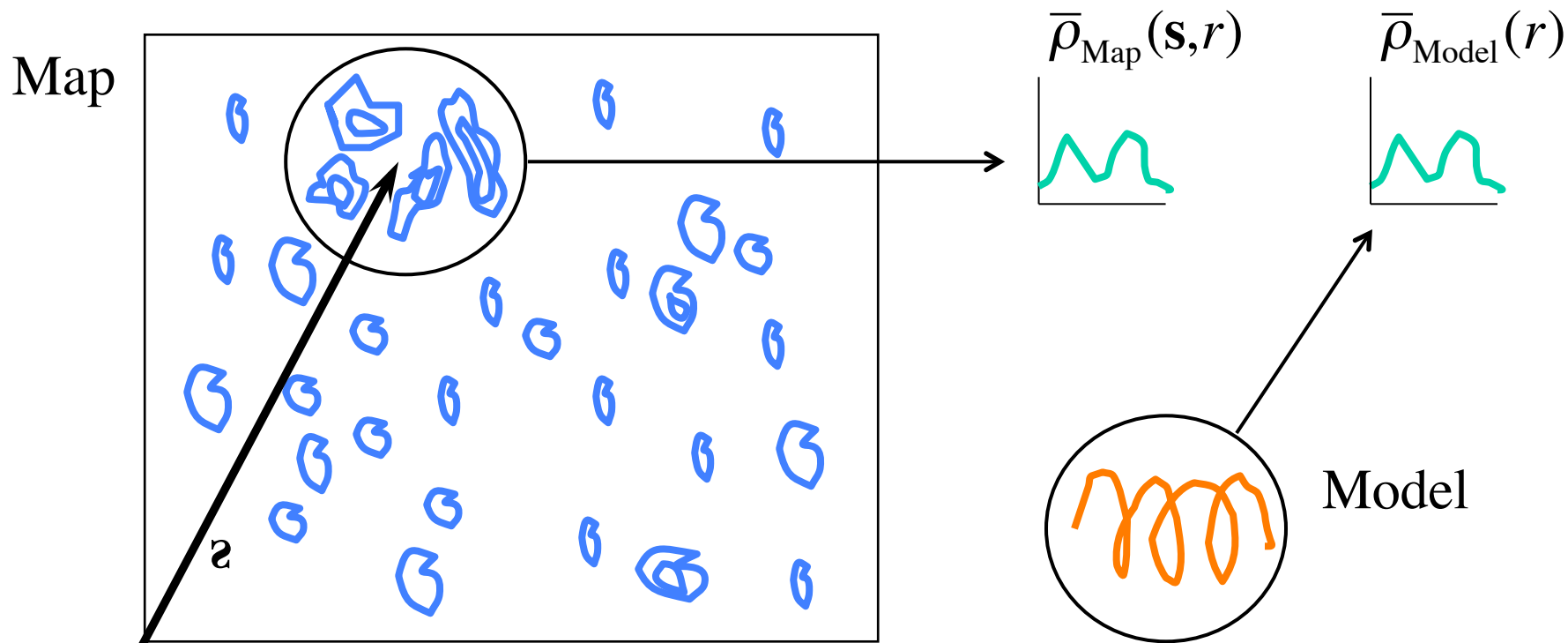
Search in the map

- Calculate 2-1 or 1-1 maps after restrained refinement of partial structure
- Flatten the **map** corresponding to the known substructure
- Calculate structure amplitudes from the modified map
- Use these **modified amplitudes** in Rotation Function
- And finally – **Phased TF**

Molrep: SAPTF

Spherically Averaged Phased Translation Function
(FFT based algorithm)

$$\text{SAPTF}(\mathbf{s}) = \int \bar{\rho}_{\text{Map}}(\mathbf{s}, r) \bar{\rho}_{\text{Model}}(r) r^2 dr$$



Molrep: Search in the map with SAPTF

1. Find approximate position:
Spherically Averaged **Phased** Translation Function
2. Find orientation:
Local Phased Rotation Function
 - Local search of the orientation in the density
3. Verify and adjust position:
Phased Translation Function

Molrep: Search in the map with SAPTF

1. Find approximate position:

Spherically Averaged **Phased** Translation Function

2. Find orientation:

Local Rotation Function

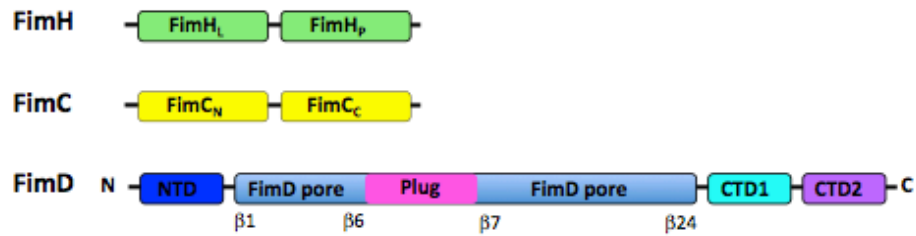
– Structure amplitudes from the density within the SAPTF sphere

3. Verify and adjust position:

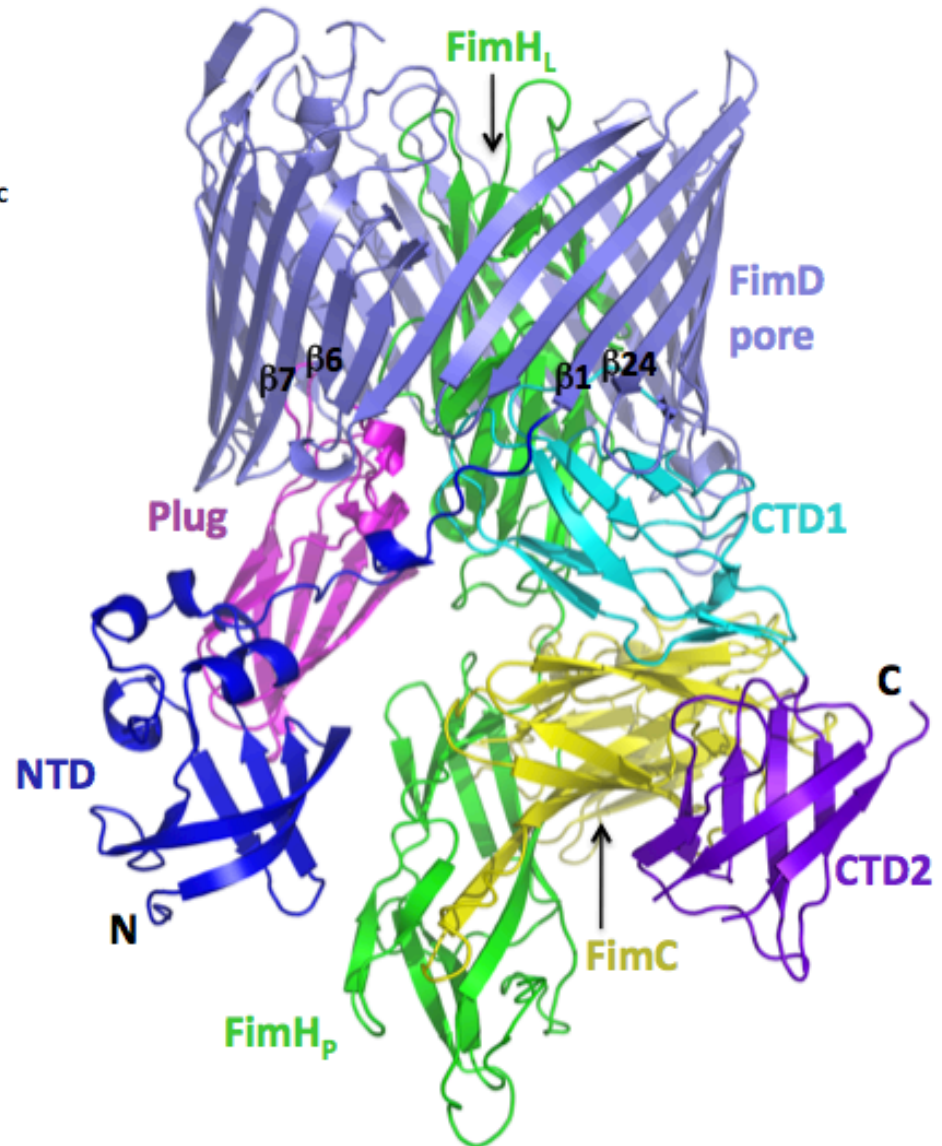
Phased Translation Function

- Local RF is less sensitive than Phased RF to inaccuracy of the model position

Example



- Asymmetric unit two copies
- Resolution 2.8 Å

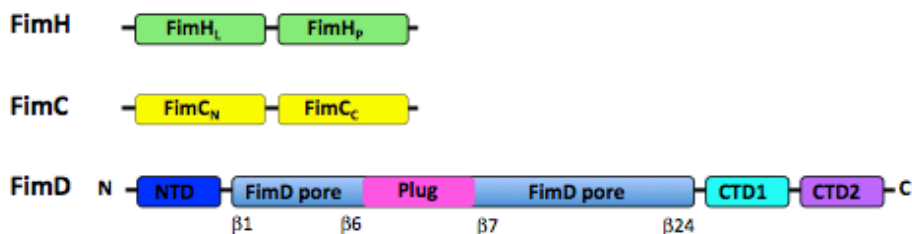


Phane et. al (2011) Nature, 474, 50-53

Usher complex structure solution

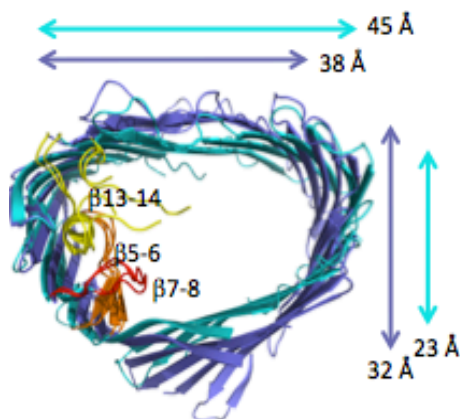
1. Conventional MR

- FimC-N + FimC-C
- FimH-L + FimH-P
- FimD-Pore



2. Jelly body refinement (Refmac)

- FimD-Pore



3. Fitting into the electron density

- FimD-Plug
- FimD-NTD
- FimD-CTD-2

4. Manual building

- FimD-CTD-1

Performance of fitting methods

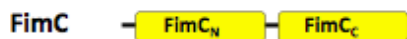
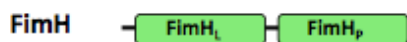
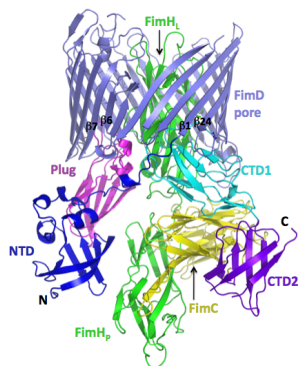


Table: the number of copies found by different methods

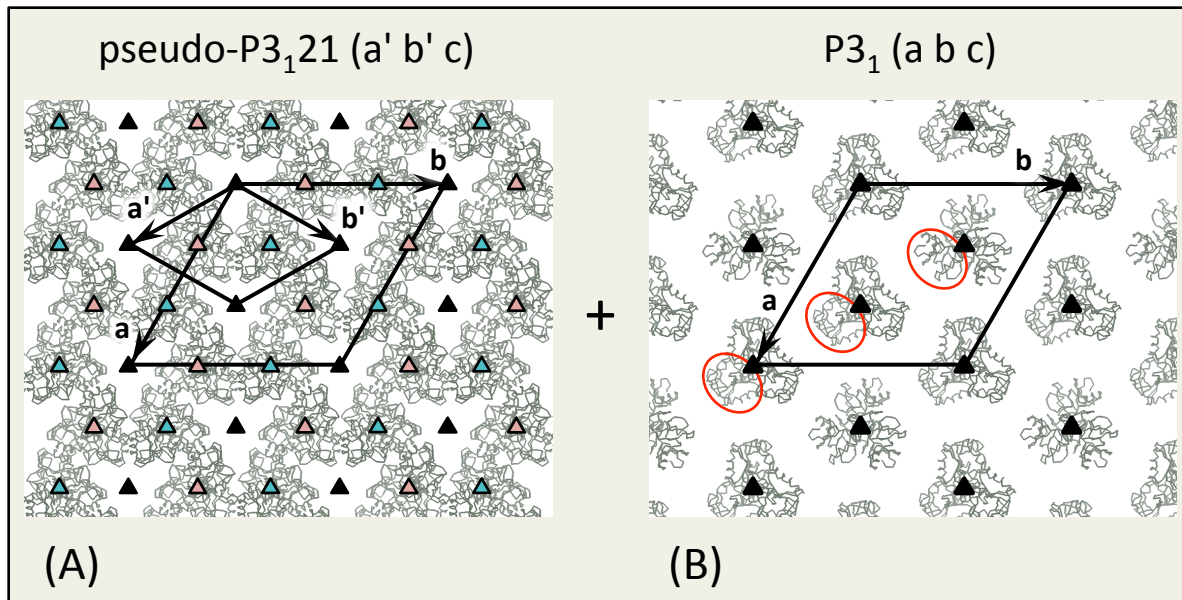
- Trying several methods is a good idea (also because of cross-validation)

	search model	sequence identity	"Masked" RF PTF	SAPTF PRF PTF	SAPTF Local RF PTF
FimD-Plug	3fip_A	38.5%	2	—	1
FimD-NTD	1ze3_D	100%	2	1	2
FimD-CTD-2	3l48_A	33.3%	—	2	—

Twinned crystal with pseudo-symmetric substructure

Human macrophage receptor CLEC5A for dengue virus

Watson, A. A. et al. (2011). J Biol Chem 286, 24208-18.

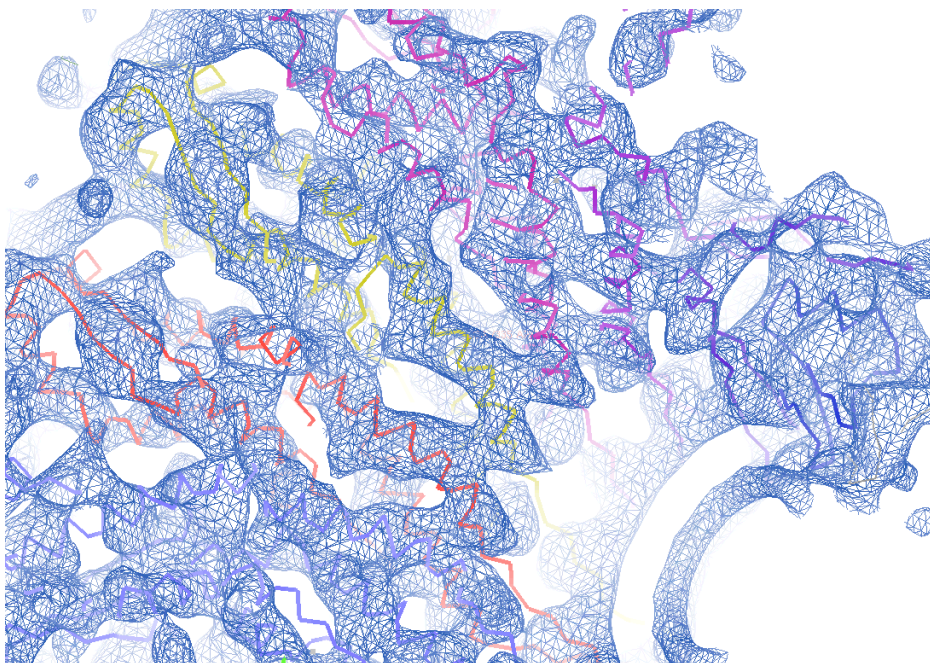


3-fold axes with respect to the true structure:

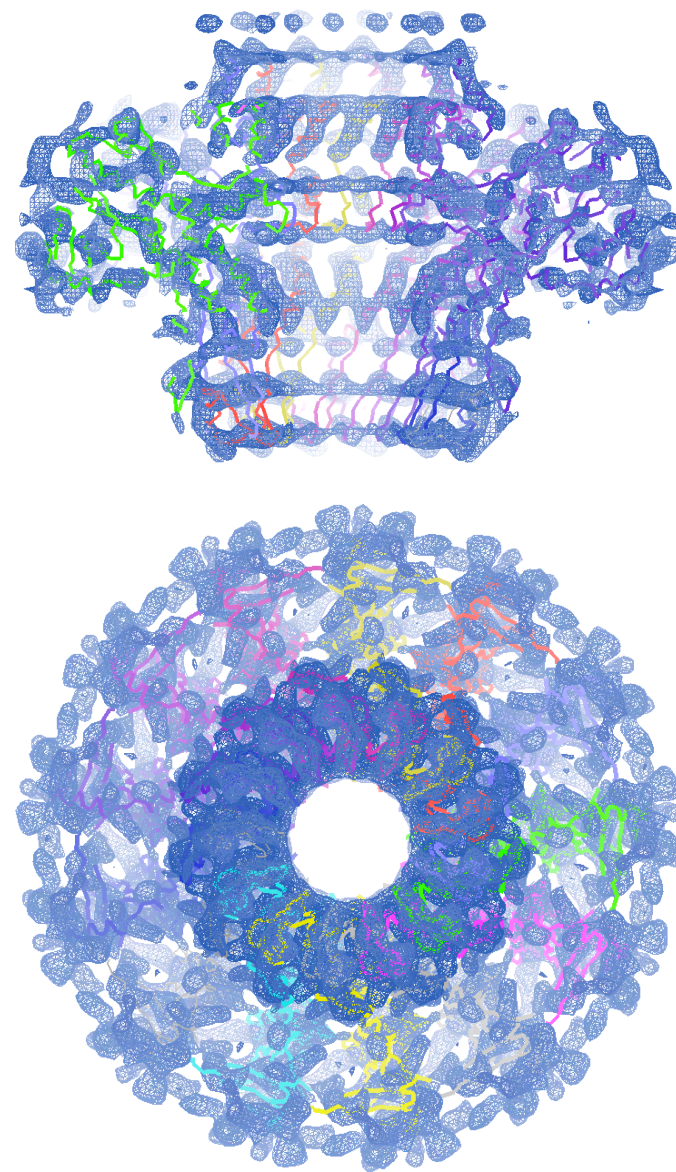
- ▲ crystallographic
- ▲▲ pseudosymmetry for (A)

- Substructure (A): Patterson search (4 copies), search in density (2 copies)
- Substructure (B): Search in the density (3 copies)

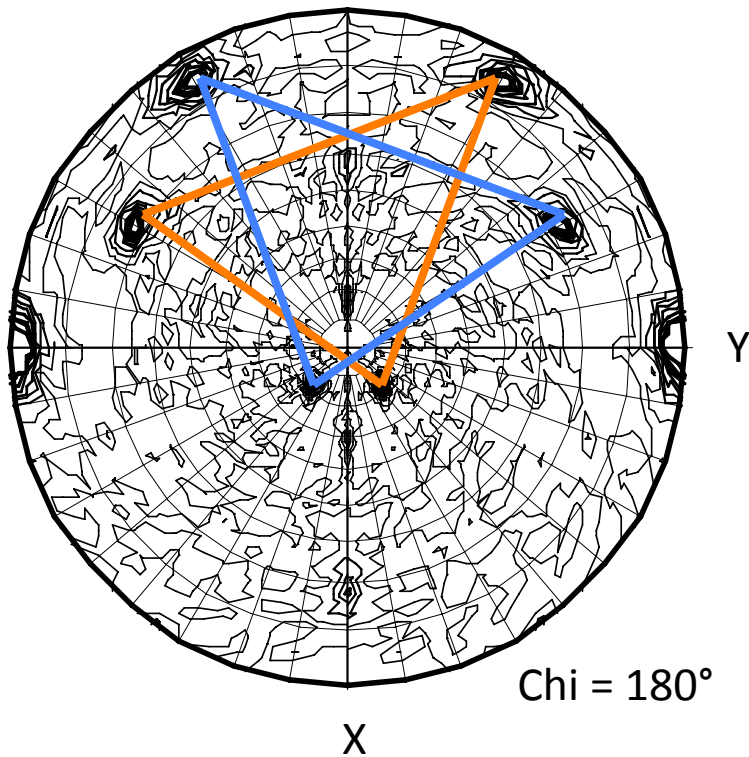
Fitting into EM maps



SPP1 portal protein



Self Rotation Function (SRF)



Preliminary analysis of X-ray data

- Oligomeric state of the protein in crystal
- Selection of oligomeric search model

Limited use

- No clear interpretation or even artifact peaks in high symmetry point groups (e.g. 622)
- different oligomers with the same symmetry

Example of SRF

- Space group P21
- One 222-tetramer in the AU

Locked Rotation Function

- Uses SRF to derive NCS operations
- Averages RF over NCS operations
- In favorable cases Improves signal to noise ratio in RF

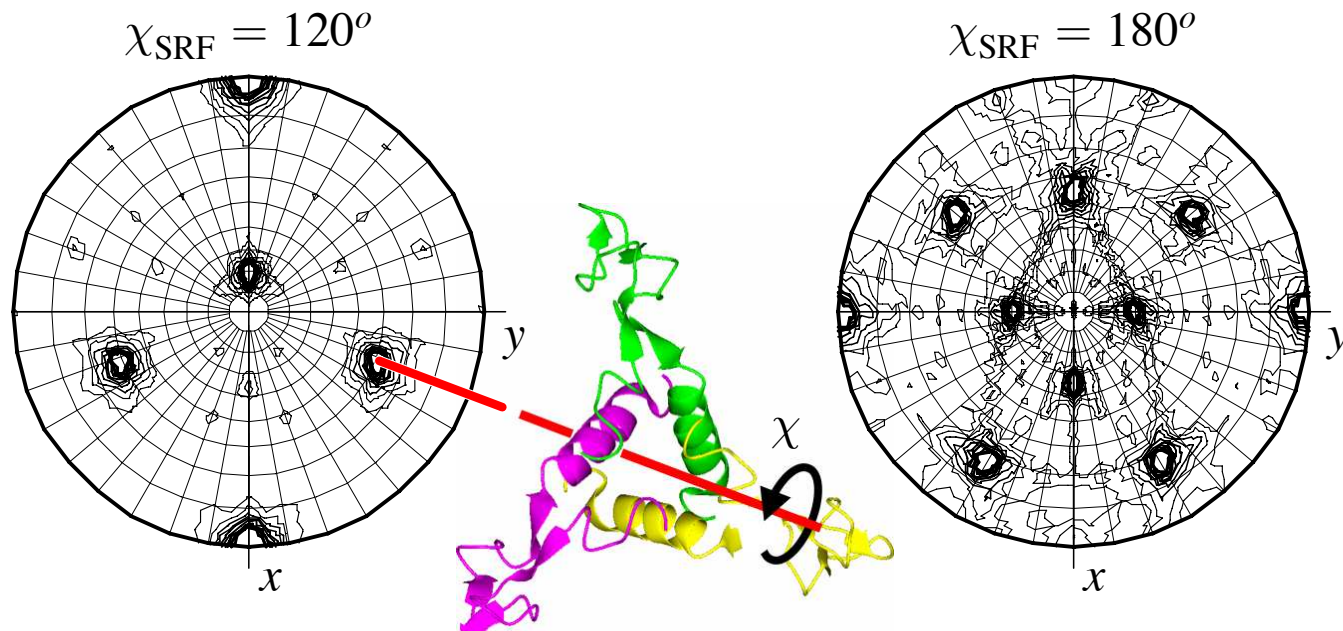
Automatic mode:

```
molrep -f s100.mtz -m monomer.pdb -s s100.seq -i <<+  
lock y  
+
```

There is an option of selecting specific SRF peaks

Also implemented in CCP4I

One-dimensional exhaustive search (exotic case)

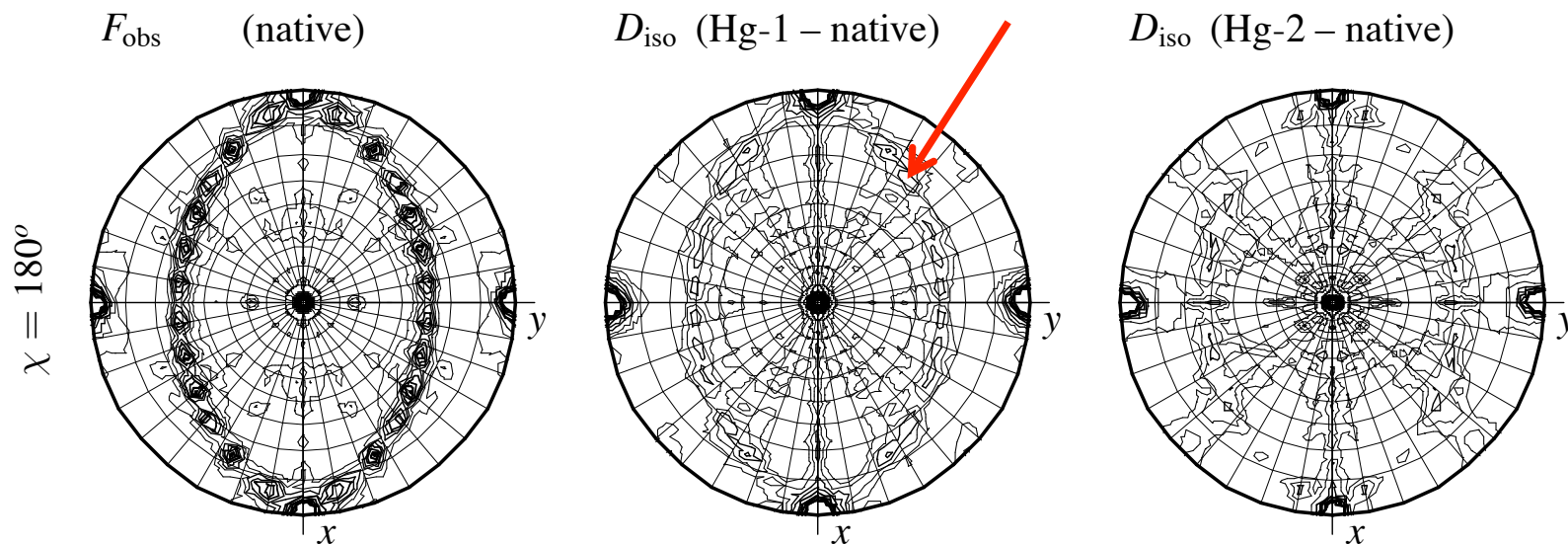


SRF helps restrict dimensionality in an exhaustive search

- Orientation of the trimer is known from the analysis of SRF
- Unknown parameter: rotation about 3-fold axis
- One-parametric exhaustive search using TF as score function

MR substructure solution (exotic case)

- Select isomorphous derivative
 - by comparing native SRF and SRF from D-iso



- Hg-substructure is a 13-atom ring (from native SRF analysis)
 - Orientation of the ring is known from the analysis of SRF
 - Unknown parameters: radius of the ring, rotation about 13-fold axis
- Two-parametric exhaustive search

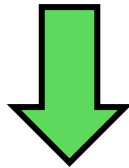
Which direction does MR go?

Automation:

- ✗ Collection of tricks
- ✓ Improvement of "standard" methods
- ✓ Better scoring system
- ✓✓ Models

Model improvement

- ✓ Structural information in MR models
- ✓ Extra information in sequence alignment
- ✓ Many homologous structures



- Remove residues that do not align
- Remove "excessive" atoms from aligned residues
- Ensemble models (several superposed models)

Single model correction:

- Chainsaw
- Molrep
- Sculptor

Preparation of ensemble models – fitting models:

- Lsqkab
- ProSMART
- SSM (also in Coot)
- Gesamt

Automated preparation of ensemble models:

- Ensembler

Molecular Replacement in CCP4

MR Programs

- AMoRe
- Molrep
- Phaser

MR Pipelines

- MrBUMP
- Balbes
- AMPLE
- Arcimboldo